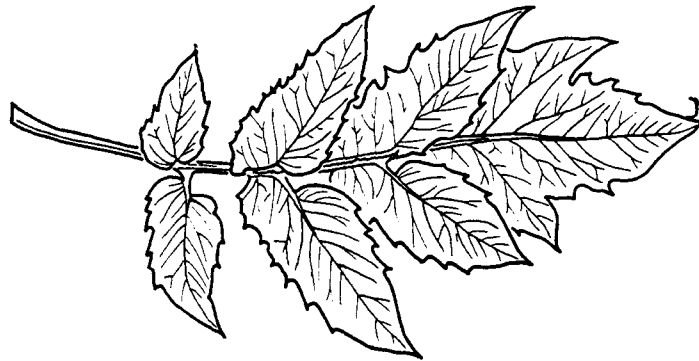


REPORT of the TOMATO GENETICS COOPERATIVE



NUMBER 12

FEBRUARY 1962

DEPARTMENT OF VEGETABLE CROPS
UNIVERSITY OF CALIFORNIA
DAVIS, CALIFORNIA

This report is a medium of exchange among members of information and stocks relating to tomato genetics. None of the information herein may be used in publications without consent of the respective authors.

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Cover design was traced from a photograph of a typical leaf of the entire (e) mutant.

FOREWORD

The Tomato Genetics Cooperative is a group of workers who have a common interest in tomato genetics and who are organized informally for the purpose of exchanging information and stocks. Participation is voluntary, and costs of activities are met by assessments to members.

TGC membership stood at 241 as of January 1, 1962, representing a slight increase over the 235 total of the preceding year. With a balance of \$298.16, TGC finances are satisfactory. This figure is not excessively large, for the majority of members are paid up for the 1961-63 triennium and expenses for 1962 and 1963 will mostly be met by this balance plus dues of new members and sale of backnumbers.

The usual annual meeting was held at Purdue University in conjunction with the AIBS meetings in August, 1961. Minutes of the meeting appear on the next page. The meeting in 1962 is again scheduled at the AIBS sessions, thanks to the cooperation of that organization. If you are planning to attend the meetings in Corvallis, Oregon in August please put the TGC meeting on your list.

New tomato mutants are being reported at such a rate that the Coordinating Committee decided to instruct the Gene List Committee to prepare the third list for this report. This new list, together with a condensed list of all previously described mutants, has been completed and appears in the following pages. We owe a special vote of thanks to Dora Hunt for taking the major responsibility for this effort and to Dick Robinson for taking over the assignment of seed stock keepers.

We are grateful to the many workers who assisted in the preparation of this Report. Thanks are due to the many members who have shared their interesting findings. We are greatly indebted to Virginia Borelli for typing the stencils on her usual high standards. (This is the ninth TGC Report for Virginia.) The skillful editing and attention to many other details by Dora Hunt are gratefully acknowledged. Thanks also to many graduate students who assisted with various details.

Four hundred copies of this Report have been issued.

Coordinating Committee

A. B. Burdick	C. M. Rick, Chairman
L. Butler	Department of Vegetable Crops
A. L. Harrison	University of California
G. B. Reynard	Davis, California

Minutes of the Purdue Meeting

August 30, 1961

The meeting took place at 12:00 noon in room 235 of the Student Union, and there was an overflow crowd present. While the orders were being taken for lunch, the chairman, L. Butler, asked each person to stand and introduce himself, and then neighbours exchanged the usual tomato gossip until after the meal had been eaten. The service was slow, and there were two tomato papers to be given at 1:00 and 1:10 in the A.S.H.S. meetings so several people had to leave right after lunch. The chairman read a report from C. M. Rick which showed that TGC has 230 members and a bank balance of \$239.49. Also that the Coordinating Committee had made the following decisions:

1. A supplementary gene list will be issued in TGC 12, and members will be canvassed to see if they will still serve as seed sources for certain mutants.
2. In response to a suggestion that TGC meet with other groups for its annual meeting, it was decided that the current practice should be continued.
3. Exchange stock lists will not be issued in TGC 12.
4. A revised membership list will be issued in TGC 12.
5. The latest revised linkage map has been transmitted to the editor of the Handbook on Growth.

The chairman then asked for discussion on the decisions. Most comments were favourable, but it was pointed out that perhaps the seed stock of mutants could be looked after by a central government agency.

A communication from T. O. Graham was read which indicated that the pedigrees for the varieties Century, Epoch, Hertford Cross, Summer Sunrise, Swift, Tecumseh, VFI 36, VFI 45, and Ware Cross would be published soon. The meeting then adjourned so that we could get back to the meetings.

V. M. Butler,

Secretary, pro tem.

LIST OF TOMATO GENES AS OF JANUARY 1962

Two previous lists of genes have been issued in TGC 4 and 9. The former was published with a list of nomenclatorial rules in Journal of Heredity 46: 22-26, 1955, and the latter with supplementary rules in the same medium 51: 167-174, 1960. During the three-year interval since the last list was issued, numerous new genes have been reported and the need for an additional rule has risen.

Recent investigations have revealed the approximate position of genes on certain chromosome arms. The following rule was therefore formulated as a means of presenting this information in the chromosome maps. It is consistent with usage in the maize and *Drosophila* groups.

1. As soon as the arm location of a gene is known, the locus numbering should be revised to reflect that information. The small arm of each chromosome is designated as the left arm, and the zero position is the left end of the small arm.

Following the procedure of the second list, two tables have been prepared. Table I lists in condensed version all genes that appeared in previous lists. Table II lists new genes that have been described since the second list was issued. The inclusion of mutants of *L. pimpinellifolium* described in Stubbe's I list (Kulturpflanzen 8:110-137, 1960) has been deferred until certain nomenclatorial problems have been settled.

Table I. CONDENSED LIST OF GENES ISSUED IN TGC 4 & 9

(Asterisk signifies genes first listed in TGC 4.)

Symbol	Name	Seed Source	Symbol	Name	Seed Source
* a ₁ (a)	anthocyaninless	Bur Rob	* ap	apetalous	Bur R
ac	apocarpous		ar	arrecta	St B
* ad	<u>Alternaria</u>	A K	* as ₁	asynaptic-1	Soost C
	resistance		* as ₂	asynaptic-2	Soost C
ae (a ₃₃₂)	entirely	Bur Rob	* as ₃	asynaptic-3	Soost C
	anthocyaninless		* as ₄	asynaptic-4	Soost C
aeg	aegrota	St R	* as ₅	asynaptic-5	Soost C
af (a ₃₂₅)	anthocyanin-free	Bur K	* as ₆	asynaptic-6	Soost C
* ag	anthocyanin	B R	* at	apricot	Jenkins R
	gainer		atn (at)	attenuata	St B
ah	Hoffman's	Rob WW	au ₁ (au, brace)	aurea	Lesley WW
	anthocyaninless				
ai (a ₃₄₂)	incomplete	Bur Rob	* aw	without	NC B
	anthocyanin			anthocyanin	
* al (a ₂)	anthocyanin	B R	* B	Beta-carotene	Tomes NC
	loser		bc (bi)	bicolor	St B
* an ¹ (an)	anantha	St	bg	bursting resistance	
* an ² (ca)	cauliflower			of fruit	
ang	angustifolia	St B	bi	bifurcate	Bur C
ao (a ₃₃₇)	anthocyanin	Bur Rob		inflorescence	
	omitted		* bk	beaked	B C

Symbol	Name	Seed Source		Symbol	Name	Seed Source	
bl	blind	R	WW	exl (ex)	exilis	St	B
bn	blunt	Brown	WW	* f	fasciated	B	Bur
* br	brachytic	B	R	fir	firma	St	B
Bt	Bursting susceptibility of fruit			* fl	fleshy	B	PAY
btl	brittle	A	Rob	fla	flavescens	St	R
bul	bullata	St	B	flav	flavida	St	B
* c	potato leaf	B	Rob	* fru ^{abb} (bu ^{abb})**	fruticosa abbreviata	B	PAY
car	carinata	St	R	fruticosa	fruticosa	St	B
* cb	cabbage			fruticosa hemiglobosa	fruticosa hemiglobosa	St	B
* Cf ₁ (Cf _{sc})	resistance to <u>Cladosporium</u> <u>fulvum-1</u>	K	A	fu	fusiformis	St	K
* Cf ₂ (Cf _{pl})	" -2	K	A	ful	fulgens	St	R
* Cf ₃ (Cf _{p2})	" -3	K	A	Fw	Furrowed	R	HWY
* Cl ₁	Cleistogamous-1	R	NC	* (G)	(interacts with r)		
* cl ₂	cleistogamous-2	R	NC	* g	grooved	C	WW
clā	clara	St	R	gf	green flesh	K	R
clau (ff)**	clausa	St	R	gh (ab)	ghost	R	Bur
cm	curly mottled	R	B	gil	gilva	St	
cn (ca)	cana	St	B	glau	glaucescens	St	K
co	cochlearis	St	B	glo	globosa	St	R
con	convalescens	St	R	gq	grotesque	R	K
cr	radial cracking resistance of fruit			gr	green ripe	K	
cs	corollaleless	K		gra	gracilis	St	K
Cu ₁ (Cu)	Curl-1	R	PAY	* gs	green stripe	R	NC
cv	curvata	St	B	* H	Hairs absent	B	R
d ₁ ^{crisp} (rob ^{crisp})	crispata	St	R	* hl	hairless	B	Bur
* d ₁ (d, rob ^{imm})	dwarf-1	B	Johnson	hp (hp ₁ , hp ₂ , bs, dr)	high pigment	Thompson	K
* d ₁ ^x (d ^x)	extreme dwarf-1	R	C	ht	hastate	R	Abdel-Al
d ₂ (rob ₂)	dwarf-2	St	NC	* I	Immunity to <u>Fusarium</u> <u>lycopersici</u>	NC	Gilbert
d ₅ (d ₂ , m-5)	dwarf-5	Bur	Rob	ig	ignava	St	K
dē	declinata	St	B	imb	imbecilla	St	R
deb	debilis	St	R	in	indiga	St	K
def	deformis	St	R	inc	incurva	St	K
deli	deliquescens	St	B	inf	infirma	St	R
dep	deprimata	St	R	int	integerrima	St	K
depa	depauperata	St	B	intro	introflexa	St	K
di	divergens	St	B	inv	invalida	St	R
dil	diluta	St	R	irr	irregularis	St	K
dim	diminuta	St	B	* j ₁ (j, lf)	jointless-1	Munger	Bur
dis	discolor	St	B	j ₂	jointless-2	Reynard	K
* dl	dialytic	R	NC	jūg	jugata	St	R
* dm (d ₂)	dwarf modifier	B	Rob	* (K)	(interacts with r)		
ds	dwarf sterile	R	Bur	* l ₁ (l, g, rub)	lutescent-1	B	R
* dv	dwarf virescent	R	C	l ₂	lutescent-2	R	K
* e (b)	entire	B	R	Lā	Lanceolate	Jenkins	R
ec	exserted carpels	PAY	K	lat	lata	St	R
* el (e)	elongate	B	Creech	* Lc	Locule reduction	PAY	NC
ele	elegans	St	R	* (lc ₁ , lc ₂ , lc ₃)	(control locule number)		
elu	eluta	St	R	* lg ₁ (lg)	light green-1	B	PAY
er	erecta			lo ₁	locule number reduced		
* ex	exserted	R	Creech				

**According to information received from Dr. Stubbe during preparation of TGC 12.

Symbol	Name	Seed	Source	Symbol	Name	Seed	Source
ls	lateral supressor	WW	R	* p	peach	B	C
luc	lucida	St	R	pal	pallida	St	R
lut	lutea	St	R	pau	pauper	Bohn	K
* m ₁ (m)	mottled-1	B	C	pc	precocious	C	Rob
M ₃ (M ₃₄₅)	Mottled-3				chromosome division		
marm	marmorata	St	R	* pe	sticky epidermis	B	PAY
* mc	macrocalyx	HWY	R	per	perviridis	St	R
md (mo, m-2)	mottled-2	Bur	R	pg ₁ (pg, pg ₃₂₉)	pale-green-1		
Me	Mouse ears	R	C			Bur Creech	
Mi	Meloidogyne incognita			Ph ₁ (Pi _{T1} , TR ₁)		Gallegly	
	resistance	Gilbert	NC		susceptibility to	A	
mn ₊ (mi _B)	minuta	St	K		Phytophthora infestans		
mo _B (I _B)	modifier of B	Tomes	NC		race T ₁		
mon	monstrosa	St	K	* pi	pistillate	R	NC
* ms ₁	male-sterile-1			pic	picta	St	NC
* ms ₂	male-sterile-2	R	C	* pr	propellor	B	R
* ms ₃	male-sterile-3	R	Bur	pro	procera	St	R
* ms ₄	male-sterile-4	R	Creech	prun	prunoidea	St	R
* ms ₅	male-sterile-5	R	C	* ps (va)	positional	B	C
* ms ₆	male-sterile-6	R	NC		sterile		
* ms ₇	male-sterile-7	R	Bur	pt	petite	Bur Creech	
* ms ₈	male-sterile-8	R	NC	pu	pulvinata	St	NC
* ms ₉	male-sterile-9	R	Bohn	pum	pumila	St	R
* ms ₁₀	male-sterile-10	R	NC	pur	purilla	St	R
* ms ₁₁	male-sterile-11	R	Creech	* r _y (r ^y , ry)	yellow flesh	R	Bohn
* ms ₁₂	male-sterile-12	R	Bur	r ₁	reddish	PAY	Bohn
* ms ₁₃	male-sterile-13	R	NC		yellow		
* ms ₁₄	male-sterile-14	R	NC	ra	rava	St	R
* ms ₁₅	male-sterile-15	R	C	* rc	rolled		
* ms ₁₆	male-sterile-16	R	Creech	re	reptans	St	NC
* ms ₁₇	male-sterile-17	R	NC	rela	relaxata	St	NC
* ms ₁₈	male-sterile-18	R	NC	rep	repens	St	NC
ms ₁₉	male-sterile-19			* ri	ridged	NC	Gilbert
ms ₂₀	male-sterile-20			rl (ra)	radial crack resistance		
ms ₂₁	male-sterile-21				of fruit		
ms ₂₂	male-sterile-22			* ro	rosette	B	C
ms ₂₃	male-sterile-23	R	Creech	ru	ruptilis	St	NC
ms ₂₄	male-sterile-24	R	Creech	rust	rustica	St	R
* mt	midget			* rv	reticulate	R	WW
mult	multiflora	St	R		virescent		
* n (nt)	nipple	B	HWY	* s	compound	B	C
na	nana	St	R		inflorescence		
* nc	narrow cotyledons	B	K	sd	sun dwarf	R	K
nd (m-4)	netted	Bur	WW	* Se	Septoria	A	Tomes
* ne ₁ (ne)	necrotic-1	B	PAY		resistance		
neg	neglecta	St	R	sf	solanifolia	R	WW
ni	nitida	St	K	sit	sitiens	St	R
not	notabilis	St	K	* sl ₁ (sl)	stamenless-1	NC	Iyall
Nr	Never ripe	R	K	sl ₂ (sl ₂)	stamenless-2	WW	
nv	netted virescent	Soost	C	sl ₃ (sl ₃)	stamenless-3		
* o	ovate	B	C	sl ₄ (sl ₄)	stamenless-4		
* (O, O', o)	(Sperical, oblate, elongate)			sl ₅ (sl ₅)	stamenless-5	WW	
* O ¹ (ol)	Oval	R		* Sm	Stemphyllium	A	Gilbert
op	opaca	St	K		resistance		

Symbol	Name	Seed Source		Symbol	Name	Seed Source	
* sp	self-pruning	B	Rob	uni	unicaulis	St	R
squa	squarrosa	St	NC	* v ₁ (v)	virescent		
* st	sterile			va ^{dec}	varia ^{decolorata}	St	R
stri	stricta	St	R	va ^{virg}	varia ^{virgata}	St	R
su	suffulta	St	NC	var	variabilis	St	R
sub	subtilis	St	R	* Ve	<u>Verticillium</u>	R	Rob
suf ^{pura}	sufflava	St	R		resistance		
sulf ^{pura} 5-100%	sulfurea ^{pura} alleles	St		vel	velutina	St	NC
sulf ^{vag} 5-100%	sulfurea ^{variegata} alleles	St		ven	venosa	St	R
Sus _a	Subsistens	St		ver	versicolor	St	NC
Sw ₁	Spotted wilt resistance-1 ^a	NC	Gilbert	* vg	vegetative	R	NC
Sw ₁ ^b	Spotted wilt resistance-1 ^b	NC	Gilbert	* vi	villous	B	K
sw ₂	spotted wilt resistance-2	NC	Gilbert	vio	violacea	St	NC
sw ₃	spotted wilt resistance-3	NC	Gilbert	vir	viridis	St	R
sw ₄	spotted wilt resistance-4	NC	Gilbert	vit	vitiosa	St	NC
sy	sunny	Bur	Bohn	* w ₁ (w)	wiry-1	B	Foskett
* t _v	tangerine	B	R	* w ₂	wiry-2 (Lindstrom)		
t	virescent	Lesley	R	* w ₃ (w ₂)	wiry-3 (Lesley)		
	tangerine			* wd	wilty dwarf	Rob	NC
tab	tabescens	St	NC	* wf	white flower	R	Bohn
ten	tenuis	St	R	* Wo ^m	Wooly	B	R
* tf (ct, tri)	trifoliolate	Jenkins	HWY	Wo ^v	Morgan's wooly	R	C
Tm ₁	Tobacco-mosaic virus resistance-1	C	NC	Wo ^v	vanWert wooly	Soost	C
Tm ₂	Tobacco-mosaic virus resistance-2	Soost	C	* wt	wilty	B	HWY
* u (u ₁)	uniform ripening	B	R	* x	modifier of I	Gilbert	NC
* ug (u ₂)	uniform grey-green	B	PAY	* Xa	Xanthophyllic	B	R
um	umbrosa	St	NC	* y (y ¹)	colorless fruit	B	R
					skin		
				yg ₁	yellow-green-1	Thompson	
				yg ₂ (yg ₂₈₂ , yc)	yellow-green-2	Bur	K
				yg ₃ (yg ₃₃₀ , ye)	yellow-green-3	Bur	Gilbert
				yg ₄ (yg ₃₃₃ , yl)	yellow-green-4	Bur	K
				* ys	yellow seedling		
				* yv	yellow virescent	R	Gilbert

Table II. LIST OF GENES JANUARY 1959 - JANUARY 1962

Gene symbol	Reference	Seed Source	Character
aba (ab)(a ₁₇₉)	Burdick, TGC-10:8	B	anthocyaninless-base. A typical green stem mutant, similar to all the others; spontaneous in San Marzano.
acu	Stubbe, 1959	St Abdel-Al	accumbens. Leaves and pinnae shortly stalked; leaf surface furrowed; older leaves strongly bent downward. Condine Red.

<u>Gene symbol</u>	<u>Reference</u>	<u>Seed Source</u>	<u>Character</u>
adp	Stubbe, 1959	St Abdel-Al	<u>adpressa</u> . Leaves epinastic; shoots, internodes and leaflets shortened; upright habit. Condine Red.
afl (af)	Rick, TGC-11:18	R Creech	<u>albifolium</u> . Seedling very slow; cotyledons white; leaves becoming normal except for gray flecks; later growth nearly normal. XL Pearson.
afr (ap)	Yu & Yeager, 1960		<u>anthocyaninless-fragile</u> . Anthocyanin absent; small, brittle, fragile plants; leaves wilt in a day. Chatham.
am (a ₃₄₀)	Burdick, TGC-10:8	B Rob	<u>anthocyanin-minus</u> . A typical green stem mutant, similar to all the others; irradiation-induced.
apl	Stubbe, 1959	St Abdel-Al	<u>applanata</u> . Leaves elongate, laxly pinnate; fruit flatter, broader, larger; spreading habit. Lukullus.
be	Lesley, TGC-8:24	Lesley Verkerk	<u>blue-green</u> . Small plants with blue-green leaves; probably an irradiation-induced translocation. Canary Export.
bip	Stubbe, 1959	St Abdel-Al	<u>bipinnata</u> . Leaves highly divided, with smaller acute segments. Lukullus.
ch	Rick, TGC-10:31	R Foskett	<u>chartreuse</u> . Greenish-yellow corolla segments. Pearson.
chln	Böhme & Scholz, TGC-11:5		<u>chloronerva</u> . Upper leaves smaller and chlorotic interveinally; strong graft response. Bonner Beste.
com	Stubbe, 1959	St Thompson	<u>complicata</u> . Plants much reduced; stems upright, stiff. Condine Red.
cor	Stubbe, 1959	St Foskett	<u>coriacea</u> . Tiny unbranched plants; all parts reduced; short leaves with plicate segments. Condine Red.
cpt	Rick, TGC-12:37	R Rob	<u>compact</u> . Highly branched compact plant; lax stems; pale, smaller leaves. XL Pearson.
Cr ₁	Butler, TGC-12:17	B	<u>Crimson-1</u> . Complementary gene effecting crimson fruit color. Philippine variety.

<u>Gene symbol</u>	<u>Reference</u>	<u>Seed Source</u>	<u>Character</u>
cr ₂	Butler, TGC-12:17	B	<u>crimson-2</u> . Complementary gene effecting crimson fruit color. Philippine variety.
Cri	Stubbe, 1959	St HWY	<u>Crispa</u> . Cri/Cri inviable without grafting; minute in all parts; leaves bright olive-green, necrotic. Cri/+ whitish brown spots on the middle of the leaflets. Condine Red.
cu ₂	Yu & Yeager, 1960		<u>curly leaf-2</u> . Small, downcurled leaves. Chatham.
cur	Stubbe, 1959	St HWY	<u>curvifolia</u> . Leaves twisted, yellow green; shortened internodes. Rheinlands Ruhm.
cva	Stubbe, 1959	St Foskett	<u>conversa</u> . Dense growth habit; older leaves lose pigment. Condine Red.
dec	Stubbe, 1959	St Thompson	<u>decumbens</u> . Leaves sparse; plant very prostrate and spreading. Lukullus.
det	Stubbe, 1959	St Verkerk	<u>detrimentosa</u> . Reduced branching; slower growth; tends to be virescent; base of young leaves tend to be yellowish. Rheinlands Ruhm.
div	Stubbe, 1959	St Rob	<u>divaricata</u> . Leaves concave; chlorotic between veins; anthocyanin in reverse. Condine Red.
dp	Yu & Yeager, 1960		<u>drooping leaf</u> . Leaf drooping, elongate, dark green; stems weak, slender, prostrate. Chatham.
fe	Stubbe, 1959	St Foskett	<u>fertilis</u> . Leaves roll ventrally; pendant. Lukullus.
flc	Stubbe, 1959	St Foskett	<u>flacca</u> . Leaves small, recurved dorsally, susceptible to over-wilting. Rheinlands Ruhm.
fr	Stubbe, 1959	St Foskett	<u>frugalis</u> . All parts reduced; plant small, compact; leaves dark green, brittle. Condine Red.
Fs	Clayberg, TGC-12:22	C Rob	<u>Fruit stripe</u> . Dark green radial stripes on immature fruits opposite and equal in number to locules; unripe fruit highly pigmented. Backcross transfer from <u>L. hirsutum</u> f. <u>glabratum</u> .
gl	Stubbe, 1959	St Verkerk	<u>glauc</u> . Leaf segments narrow, acute; virescent at growing point. Condine Red.

<u>Gene symbol</u>	<u>Reference</u>	<u>Seed Source</u>	<u>Character</u>
glb	Stubbe, 1959	St Verkerk	<u>globularis</u> . Internodes and leaves shortened; leaves densely pinnate, yellow tinged. Rheinlands Ruhm.
glc	Stubbe, 1959	St Verkerk	<u>glaucophylla</u> . Leaves light yellow or gray-green, acuminate. Rheinlands Ruhm.
gri	Stubbe, 1959	St Thompson	<u>griseifolia</u> . Leaves small, strongly recurved dorsally, gray-green; trichomes longer. Rheinlands Ruhm.
he	Stubbe, 1959	St Foskett	<u>heteroidea</u> . Weak plant; internodes shortened; leaflets somewhat narrowed; older leaves glossy, brittle, pale green. Condine Red.
Hrt	Rick, TGC-12:38	R	<u>Hirtum</u> . Increased density of large trichomes; incompletely dominant. Peruvian tomato var.
ic	Stubbe, 1959	St Thompson	<u>inclinata</u> . Internodes and leaves shortened; older leaves epinastic; plant prostrate. Rheinlands Ruhm.
id	Stubbe, 1959	St HWY	<u>indehiscens</u> . Broad leaf segments; connate sepals; fruit irregularly cracked. Rheinlands Ruhm.
imp ^{dia}	Stubbe, 1959	St Foskett	<u>impedita</u> ^{distincta} . Plants short, weak; leaves chlorotic at margins; plants smaller than <u>imp</u> ^{eg} . Condine Red.
imp ^{eg}	Stubbe, 1959	St Foskett	<u>impedita</u> ^{exigua} . Like <u>imp</u> ^{dia} but plant larger. Condine Red.
j ₂ ⁱⁿ	Joubert, TGC-11:13	Joubert R	<u>jointless-2</u> with <u>incomplete</u> action. Pedicel joints with normal form but do not separate when fruits ripen. ex. Riverside x Sunneva.
lae	Stubbe, 1959	St Foskett	<u>laesa</u> . Axillary sprouts later necrotic; primary leaves dainty. Rheinlands Ruhm.
lg ₂ (1i)(pg ₃₉₂)	Burdick, TGC-10:9	B Verkerk	<u>light green-2</u> . Cotyledons light yellow (like <u>ye</u>); leaves pale green; leaf shape normal; classification sometimes difficult; irradiation-induced in seed of Red Cherry inbred.
lg ₃ (1t)(pg ₄₈₃)	Burdick, TGC-10:9	B Verkerk	<u>light green-3</u> . Cotyledons and leaves light green; cotyledons fade to yellow; mature plant pale green; irradiation-induced in seed of Red Cherry inbred.

<u>Gene symbol</u>	<u>Reference</u>	<u>Seed Source</u>	<u>Character</u>
lg ₄	Burdick, TGC-11:6	B Verkerk	<u>light green-4</u> . Yellow green cotyledons, light green leaves; seedling classification dependable; 10,000r X-ray-induced in pre-soaked seed of Red Cherry inbred.
lg ₅	Kerr, TGC-12:30	K Verkerk	<u>light green-5</u> . Light-green foliage color. PI 126,954 <u>L. pimpinellifolium</u> .
lu	Stubbe, 1959	St	<u>luteola</u> . Dark yellow flowers. <u>Lukullus</u> .
ma	Stubbe, 1959	St	<u>macrocarpa</u> . Leaves distinctly lengthened; fruit large and smooth. <u>Lukullus</u> .
mar	Stubbe, 1959	St	<u>marcescens</u> . Plants small, erect; leaves dark green, lightly wilted. <u>Lukullus</u> .
mnx (min ^{ex})	Persson TGC 10:27-28	Persson	<u>extreme miniature</u> . Severely stunted chlorina mutant with 1 cm. internodes; flowers only after grafting on +.
ms ₂₅	Rick, TGC-10:35-37	R Rob	<u>male-sterile-25</u> . Stamens slender, fore-shortened, pale; stigmas mostly exposed. Red Top.
ms ₂₆	Rick, TGC-10:35-37	R Rob	<u>male-sterile-26</u> . Stamens variable, greatly distorted; stigmas 100% exposed. Van's Early.
ms ₂₇	Rick, TGC-10:35-37	R Rob	<u>male-sterile-27</u> . Stamens slender, diverging distally, pale; stigmas mostly not exposed. Van's Early.
ms ₂₈	Rick, TGC-10:35-37	R Rob	<u>male-sterile-28</u> . Stamens very slender, slightly shorter, pale; stigmas mostly exposed. XL Pearson.
ms ₂₉	Rick, TGC-10:35-37	R Rob	<u>male-sterile-29</u> . Stamens slender, often separate below, pale, often greenish; stigmas mostly exposed. CPC-2.
ms ₃₀	Rick, TGC-10:35-37	R Rob	<u>male-sterile-30</u> . Stamens highly modified, very slender, free, twisted, pale or greenish yellow; stigmas 100% exposed. San Marzano.
ms ₃₁	Rick, TGC-10:35-37	R Rob	<u>male-sterile-31</u> . Stamens smaller, very pale; stigmas 100% exposed. VF-6 Pearson.
ms ₃₂	Rick, TGC-10:35-37	R Rob	<u>male-sterile-32</u> . Stamens greatly reduced and shrunken; very pale, often brown; stigmas 100% exposed. Primitive cultigen from Colombia.

<u>Gene symbol</u>	<u>Reference</u>	<u>Seed Source</u>	<u>Character</u>
ms ₃₃	Rick, TGC-12:39	R	<u>male-sterile-33</u> . Anthers very irregular, yellow-green; no pollen; breakdown in early meiosis; stigmas exposed; VF-11 Pearson.
ms ₃₄	Rick, TGC-12:39	R	<u>male-sterile-34</u> . Anthers short, pale; no pollen; meiosis normal, possibly delayed; stigmas exposed. VF-11 Pearson.
ms ₃₅	Rick, TGC-12:39	R	<u>male-sterile-35</u> . Anthers small, slender, pale; empty PMC in locules; breakdown in different stages of meiosis; stigmas exposed. A-1.
mu	Stubbe, 1959	St	<u>multinervis</u> . Internodes and leaves somewhat shortened; foliage full pale green, veins normal green. Condine Red.
muf	Stubbe, 1959	St	<u>multifolia</u> . Leaves elongate, highly divided, gray-green; interveinal areas blistered. Rheinlands Ruhm.
ne ₂	Yu & Yeager, 1960		<u>necrotic-2</u> . Leaves with many grayish necrotic spots; plant small, weak, killed by defoliation. Chatham.
Nec	Martin, 1961	Martin	<u>Necrotic</u> . Hypertrophic pustules along veins of leaves; followed by chlorosis, then necrosis. Backcross transfer from <u>L. chilense</u> .
ob	Stubbe, 1959	St	<u>obscura</u> . Small plants with reduced branching; early foliage dark green. Rheinlands Ruhm.
oc	Stubbe, 1959	St	<u>ochroleuca</u> . Leaves discolored, becoming white variegated. Exposed unripe fruit yellowish gray. Rheinlands Ruhm.
Od	Rick & Dempsey, TGC-10:37	R Bohn	<u>Odorless</u> . Herbage has little or no characteristic tomato odor. Galápagos accession. Primitive cultivated var.
og	Rick & Dempsey, TGC-11:21	R Gilbert	<u>old gold</u> . Corolla tawny orange. Mutant in <u>L. chilense</u> backcrossed to var. Pearson.
oli	Stubbe, 1959	St	<u>olivacea</u> . Leaves crinkled, malformed, spotted tan basally. Rheinlands Ruhm.

<u>Gene symbol</u>	<u>Reference</u>	<u>Seed Source</u>	<u>Character</u>
pen	Stubbe, 1959	St Abdel-Al	<u>pendens</u> . Leaves broad, convex, pendant, glossy. Condine Red.
pg ₂ (pa) (pg ₃₉₀)	Burdick, TGC-10:8	B Creech	<u>pale green-2</u> . Cotyledons and leaves pale green; phenotype good; irradiation-induced in seed of Red Cherry inbred.
pg ₃ (pl) (pg ₃₉₁)	Burdick, TGC-10:8	B Creech	<u>pale green-3</u> . Cotyledons pale gray-green and leaves seem to fade to a dull yellow color; easy classification; irradiation-induced in seed of Red Cherry inbred.
pla	Stubbe, 1959	St Verkerk	<u>plana</u> . Leaves gray-green with yellowish speckling. Condine Red.
pli	Stubbe, 1959	St Rob	<u>plicata</u> . Plant reduced; leaves plicate, narrow; strong anthocyanin. Lukullus.
pol	Stubbe, 1959	St Thompson	<u>polylopha</u> . Short internodes; leaves plicate and twisted; fruit flat, fasciated. Lukullus.
prec	Stubbe, 1959	St Thompson	<u>procumbens</u> . Internodes and leaves shortened; leaves blistered and lightly mottled yellow. Condine Red.
rig	Stubbe, 1959	St Verkerk	<u>rigida</u> . Leaves small, rigid, never pendant, yellowish green. Condine Red.
rot	Stubbe, 1959	St HWY	<u>rotundifolia</u> . Short internode; leaves short, broad, blistered. Rheinlands Ruhm.
Rs	Yu & Yeager, 1960		<u>Suppressed root</u> . Greatly restricted root development. Chatham.
sem	Stubbe, 1959	St HWY	<u>semiglobosa</u> . Small compact plant; very short internodes; small dark leaves. Condine Red.
si	Stubbe, 1959	St Foskett	<u>sinuata</u> . Reduced growth; leaves small, wavy, yellow-green. Rheinlands Ruhm.
sn	Lesley & Lesley, TGC-11:15		<u>singed</u> . Epidermal hairs smaller; trichomes crooked, curved, and otherwise distorted. Canary Export.
spa	Stubbe, 1959	St Foskett	<u>sparsa</u> . Small weak plants, leaves emerge yellow-green, later blotched whitish yellow-green. Condine Red.
sph	Stubbe, 1959	St Rob	<u>sphaerica</u> . Very short spherical plant; older leaves small, dark green. Condine Red.

<u>Gene symbol</u>	<u>Reference</u>	<u>Seed Source</u>	<u>Character</u>
sr (sm)	Yu & Yeager, 1960		<u>slender stem</u> . Stem slender, stiff, upright, with few branches; leaves elongate, blue-green, with few segments. Chatham.
ste	Stubbe, 1959	St Rob	<u>sterilis</u> . Leaves very densely pinnate, leaflets reduced, weakly pale green; many flower buds abort; low normal fruit set. Condine Red.
sua	Stubbe, 1959	St Verkerk	<u>suffusa</u> . Leaves broad, paler, prematurely yellowing. Rheinlands Ruhm.
suc	Stubbe, 1959	St Foskett	<u>succedanea</u> . Small delicate plants; leaves small, broad, dull green, prematurely yellowing. Condine Red.
ta	Stubbe, 1959	St Foskett	<u>tarda</u> . Small dainty plant; leaves reduced, glossy, gray-green. Condine Red.
tl	Langridge & Brock, 1961	Brock Rob	<u>thiaminless</u> . Cotyledons normal; leaves small, yellow, developing green veins, dying prematurely; lethal; viable and normal if fed thiamin.
to	Stubbe, 1959	St HWY	<u>torosa</u> . Reduced branching; determinate growth; flowers fasciated. Condine Red.
tp	Yu & Yeager, 1960		<u>tripinnate leaf</u> . Leaves tripinnately compound; retarded growth. Chatham.
tr ₁ (tr)	Stubbe, 1959	St HWY	<u>truncata</u> . Small weak plants; leaves scanty, gray-green. Condine Red.
v ₂	Kerr, TGC-12:30	K Rob	<u>virescent-2</u> . New leaves pale under greenhouse conditions; frequently undistinguishable in field. Breeding line.
w ₄	Rick, TGC-11:18	R Foskett	<u>wiry-4</u> . Similar to previously-described wiry mutants; leaves progressively more reduced. Pearson.
wv	Rick, TGC-10:32	R Creech	<u>white virescent</u> . Plant retarded; cotyledons whitish yellow; leaves start chlorotic, always with white speckling. Peruvian domestic variety.

<u>Gene symbol</u>	<u>Reference</u>	<u>Seed Source</u>	<u>Character</u>
yg ₅ (yw) (yg ₃₈₈)	Burdick, TGC-10:8	Bur HWY	<u>yellow-green-5</u> . Very yellow cotyledons and leaves; slow growing; good plant viability; irradiation-induced in Red Cherry inbred by seed treatment; the most yellow mutant we have.
yg ₆ (yo) (yg ₃₈₉)	Burdick, TGC-10:8	Bur HWY	<u>yellow-green-6</u> . Bright yellow cotyledons and leaves; seedling etiolates; plant very vigorous; gametic and zygotic viability good; irradiation-induced in seed of Red Cherry inbred.
yv ^{ms}	Hagemann, TGC-12:27	St	<u>yellow-virescent</u> ^{masculosterilis} . Male-sterile allele of yv; seedling yellow becoming yellow green (field) or light green (greenhouse). Condine Red.
yv ^{mut}	Hagemann, TGC-12:27	St	<u>yellow-virescent</u> ^{mutabilis} . Male-sterile variegated allele of yv; green color frequently mutating to yv ^{ms} . Condine Red.
(no symbol)	Gröber, TGC-11:12	lethal s	<u>lethal seedling?</u> Dies in seedling stage at low temperatures; survives and is nearly normal if grown above 26°C or grafted on +. Condine Red.

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KEY TO STOCK SOURCES IN TABLES I AND II

A	C.F. Andrus	K	E.A. Kerr	Rob	R.W. Robinson
B	L. Butler	NC	North Central Regional	St	H. Stubbe
Bur	A.B. Burdick		Station U.S.D.A.	WW	W. Williams
C	C.D. Clayberg		Ames, Iowa	HWY	H.W. Young
		R	C.M. Rick	PAY	P.A. Young

Gene List Committee

C. D. Clayberg, Chairman
 L. Butler P. A. Young
 R. W. Robinson C. M. Rick
 Dora G. Hunt

PART IRESEARCH NOTES

Bills, R. W., and M. W. Martin

Embryo sacs in hybrids between

L. esculentum and S. lycopersicoides.

The study reported herein was conducted to determine the

feasibility of using a hybrid

between Lycopersicon esculentum

and Solanum lycopersicoides as a bridge for transferring into Lycopersicon the genes which control the apparent immunity from curly top virus infection that is expressed by certain species of Solanum.

Cuttings of the F₁ hybrid used in the study were obtained from C. M. Rick, who made the cross. The commercial tomato variety V. R. Moscow was used in reciprocal crosses with the hybrid and as the standard of comparison for the studies on the embryo sac and pollen-tube growth. Fruits for study were obtained from the following pollinations:

- (a) selfing the hybrid
- (b) crossing hybrid x V. R. Moscow
- (c) crossing V. R. Moscow x hybrid
- (d) no pollination of the hybrid or V. R. Moscow
- (e) selfing of V. R. Moscow

To the pedicel of approximately half of the flowers in each of the pollination groups tested, the growth hormone parachlorophenoxyacetic acid at a concentration of 60 ppm was applied from immediately after pollination to 24 hours after. The other half of the flowers were left untreated. In all cases in the first four pollination groups those receiving no hormone treatment abscised; only those flowers receiving hormone treatment remained on the plant and developed fruit.

The intergeneric hybrid normally produces numerous small parthenocarpic fruits, but occasionally unusually large fruits which show some seed development are produced. The embryo sacs in these large naturally-occurring fruits were also studied.

Fruits were taken at intervals from the time of anthesis up to 30 days after pollination. In the case of the large naturally-occurring fruits, both young and mature fruits were collected. The fruits were killed and fixed, and the pericarps were removed. The locular contents were prepared for sectioning by the paraffin method. The sections were mounted serially and stained in safranin and fast green.

The development of the embryo sac and embryo in V. R. Moscow followed closely the pattern described by Cooper (1931, Am. J. Bot. 18:739-748) and Smith (1935, Cornell Univ. Ag. Exp. Sta. Mem. 184). In no case was embryo development observed in the hybrid pollinated with its own pollen or with the pollen of V. R. Moscow nor was embryo development observed in V. R. Moscow pollinated by the hybrid pollen. At anthesis the hybrid showed very few well-developed embryo sacs and those observed were not complete in their development. Within 8-10 days after pollination the embryo sac in the intergeneric hybrid was filled with a proliferation of cells. At ovule maturity this embryo sac area appeared as a brown area of dead oxidized cells in the aborted, nonviable seeds. The pattern of development follows that described

as the collapsed type by Rick (1946, Am. J. Bot. 33(4):250-256) in his work on sterile-ovule development in Lycopersicon esculentum.

A study also was made to determine whether any stigma or stylar incompatibilities were present in the Lycopersicon-Solanum hybrid. Stigmas and styles were collected from V. R. Moscow and the Lycopersicon-Solanum hybrid at 24-hour intervals from the time of pollination up to 72 hours after pollination. The hybrid pollen did not germinate either on its own stigma or on that of V. R. Moscow. The V. R. Moscow pollen germinated on both its own stigma and that of the hybrid. Pollen tubes of V. R. Moscow were observed throughout the length of the style of the hybrid.

Pollen from the intergeneric hybrid was stained with acetocarmine to determine by stainability the percentage of viable-appearing pollen grains produced by the hybrid. The results of this study were the same as the results reported by Rick (1951, P.N.A.S. 37:741-744). About 0.5 percent of the 1000 pollen grains counted stained darkly. Because of the small number of apparently-normal pollen grains from the hybrid, the chance of finding in vivo germination was rather small, even if it is assumed that all dark-staining pollen grains are viable. Therefore, even though observation of 70 stigmas pollinated with hybrid pollen failed to disclose any germination, there is still a question whether germination of hybrid pollen does occur on either its own stigma or that of V. R. Moscow.

The results of this study indicate that even though germination and tube growth of V. R. Moscow pollen occur normally on the hybrid pistil, fertilization and subsequent embryo development do not occur. Failure of fertilization is probably due to a lack of viable ovules which in turn is the result of the irregular meiotic behavior of the intergeneric hybrid. In pollen formation on the Lycopersicon-Solanum hybrid only a few normal-appearing pollen grains are produced and the same pattern would be expected in the formation of ovules.

Embryo sacs collapse in the Lycopersicon-Solanum hybrid at all stages of development from the megaspore to the mature embryo sac. Hypertrophy of the integumentary cells and proliferation of the cells within the embryo sac accompany the collapse.

The above results indicate that the chances of achieving successful crosses or selfs on the hybrid are quite small.

Boynton, J. E. The use of blenders
to facilitate seed extraction.

Recently the Waring Blendor has
been successfully used to
eliminate many of the laborious

hand operations involved in tomato seed extraction. A model No. P B-5A Waring Blendor coupled to a Powerstat variable transformer model 3PF 116, which serves as a convenient speed reduction device, were employed.

Use of the blender to macerate whole tomatoes, followed by fermentation of the blended pulp, has proved to be a convenient and rapid method of seed extraction for moderate to large numbers of fruits. The blender jar is loaded with whole tomatoes, sufficient water added to cover the revolving blades, the Powerstat set at 60-80 volts, and the blending action allowed to continue very briefly, 30-45 seconds--just sufficient to chew the fruit into coarse segments releasing the seeds. The blended pulp, skins, and free seeds are then poured into cans and allowed to ferment for approximately two days--just long enough for the gelatinous matrix around the seed to break down. As the ferment is decanted, the skin and pulp rise to the top, while the seeds settle to the bottom in the usual fashion.

The glass blender jars of this model are adequate for fruit lots of as many as 12 moderate-size fruit or fewer of larger-size fruit. A larger Waring Blendor, model CB4, with a 4-quart stainless steel jar, is available and should have sufficient capacity to handle most large fruit lots.

An alternative procedure for use of the Waring Blendor for immediate extraction of seeds, without fermentation, appears to be practical in certain instances--particularly with very small-fruited L. esculentum types and with fruits of the wild species. Whole tomatoes are placed in the blender jar with sufficient water to cover the revolving blades. The voltage regulator of the Powerstat is set initially at 50-80 volts in order to break up the whole fruit. As soon as this is accomplished, the voltage is reduced to + 30V, just sufficient to keep the blender blades in motion, and the blender allowed to run for 2-5 minutes, depending upon size and ripeness.

Small volumes of blended pulp can be decanted in the blender jar itself, or larger volumes transferred to another container. If an appropriate time and voltage combination is employed, the seeds will rapidly settle to the bottom, free of their gelatinous matrix. Since the skin segments usually float to the surface, they can be poured off with the finely blended pulp.

No significant damage to mature seeds--either visible effects or impairment of subsequent germination--results from this extended treatment provided a sufficiently low voltage (30-40 volts) is used. If several blender jars are used one can be loaded, another decanted, while a third lot of fruit is being blended.

A modification of the immediate extraction procedure has been found useful, particularly with the extremely large-fruited L. esculentum varieties, in which the separation of the skin segments from the seeds poses a problem. The tops of the whole tomatoes are sliced off and the entire contents--core, locules, and contents--are scooped out, using a tablespoon with sharpened edges. The foregoing procedure is then followed except that a slightly lower initial voltage is required to start the blender.

Butler, L. Crimson,
a new fruit colour.

In 1953 we obtained seed of a Philippine variety which had fruits which were a distasteful

muddy purple colour. Since this fruit colour looked like another good genetic factor, crosses were made to three tester stocks, and the following results were obtained:

P_1	X	P_1	F_1	F_2
Phil.	X	825	red	155 red: 47 crim.: 55 muddy: 3 muddy crim.
Phil.	X	902	red	50 red: 16 crim.: 6 muddy: 4 muddy crim.: 14 yellow
Phil.	X	e nc	crim.	55 red: 130 crim.: 40 muddy: 20 muddy crim.

It is obvious that muddy is a recessive, and the F_2 ratios show that it gives good monogenic segregation, and its linkage relationship will be discussed in another note. The fruit colour of the F_1 's of the third cross was entirely unexpected, the fruits were bright red and the Hunter was 134 as compared with Improved Garden States 116 and John Baer 90. Many of these F_1 fruits were normal sized but seedless, indicating that the fruits will set under poor conditions (winter greenhouse). This lack of seeds may partly account for the very high Hunter reading, but the different shade of red was quite apparent and was easily classified in all three F_2 's. From these data, providing we are willing to accept the proposition that muddy will conceal some of the

crimsons, the most simple explanation is the interaction of two genes, one dominant and the other recessive. This would give the following genotypes for the first two crosses:

$\underline{Cr_1 Cr_1} \underline{cr_2 cr_2} \times \underline{cr_1 cr_1} \underline{Cr_2 Cr_2}$ giving a red F_1 and 13 red to 3 crimson in the F_2 , and the data fit this hypothesis. For the third cross the genotypes would be: $\underline{Cr_1 Cr_1} \underline{cr_2 cr_2} \times \underline{cr_1 cr_1} \underline{cr_2 cr_2}$ giving crimson F_1 and 3 crimson to 1 red in the F_2 .

Following these observations the seed stocks were turned over to a student at one of the experimental farms, and in spite of the lapse of time, the above hypothesis has never been fully tested.

Butler, L. The cross-over values for chromosome 2.

	m	d	p	op	suf	dil	aw	ps	o	ro	Me	Cu	wv	bk	Wo	ms ₁₀	s
dv	6.8	5.2	15.0				18.9	15.5	28.0	20.1		20.3		36.1	29.1	14.6	34.2
m		2.4	7.2	6.5			14.9		28.0	5.9	26.0	23.0		20.5	29.2	35.7	22.0
d			3.0	13.5	14.0	14.4	18.2	11.8	15.8	23.4	25.0	27.0	27.8	27.9	33.2	30.0	31.6
p					7.3	15.2	14.8	10.5	14.3					26.0	26.8	16.3	28.1
op							28.7				12.7				17.6		
suf								5.5		11.0	5.9		15.5		33.1	25.5	29.0
dil									11.0		7.3	7.0	5.8		16.0	5.7	12.4
aw																	
ps																	
o																	
ro																	
Me																	
Cu																	
wv																	
bk																	
Wo																	
ms ₁₀																	
s																	

Backcrosses are being made and the chromosomes are being looked over for deficiencies in order to eliminate some of the inconsistencies which show up in the above values.

Butler, L. Muddy fruit,
a phenotype of the gene gf.

The dirty brownish purple colour
of the fruit of the Philippine
variety was at first thought to

be the expression of a new gene, and since the fruit in its early stages contained a lot of anthocyanin pigment, we thought this colour was the result of anthocyanins. Chemical analysis proved that it was not an anthocyanin, but probably a breakdown product of chlorophyll. In yellow-fruited segregates, the muddy colour did not develop, although the unripe fruits were often purple; instead the ripe fruits appeared to be similar to the ripe fruits of Staygreen caused by the gf gene. No crosses were made to check allelism but a large backcross made for another purpose shows close linkage with al. The data are:

Al red 36: Al muddy 856: al red 880: al muddy 34 or a cross-over of 3.8%.

This is much closer than the map shows but until a test is made for allelism it is better to consider muddy as the expression of the gf and R genes with the possible addition of an anthocyanin intensifier. It is interesting to note that 43 of the 58 muddy segregates had green seed, indicating pleiotropic effect or linkage.

Butler, L. The linkage
of divergens di.

The seed of this mutant was
obtained from Stubbe, and while
it is named for its habit of

growth, it was found that the seedlings could be classified by their white stems. In many cases there is a deficiency of di plants, the data obtained so far being:

Test gene	++	+ t	di +	di t	chi square t	di	linkage
d ₁	906	355	225	34	0.3	51.3	20.4
br	816	259	164	24	4.5	68.9	4.9
c	585	205	202	51	0.4	0.3	0.3
a	270	108	106	26	0.4	0.2	4.1
l	123	33	15	11	0.1	11.1	3.7
e	745	382	318	9	10.1	4.9	124.0**
Xa	266	83	68	40	8.5	0.2	2.4
au	1086	272	300	73	23.3	11.1	0.0
ao	364	133	131	26	0.3	0.3	6.4
tf	2255	801	631	226	2.0	0.3	0.0
ah	2474	804	748	240	0.6	7.7	0.0
rv	181	50	60	10	4.1	0.5	1.3
sl ₁	97	36	41	0	1.8	0.2	11.5*

These data show that e and di are linked so this gene can be placed on chromosome 4. It also appears that sl₁ is probably linked with di and may also be on this chromosome, and a three point backcross is being prepared to check on this.

Butler, L. The tomato linkage
map and the chromosomes.

Up to the present time the
mapping of the linkage groups
has proceeded on the proposition

that the chromosomes are stable, and that the tomato is relatively free from inversions, deficiencies, and translocations. The map has also been built on the supposition that cross-overs are more or less equally frequent in all regions of the chromosome. Neither of these propositions seems to hold. Our studies of the chromosomes reveal many abnormalities in the standard tester

stocks, and the linkage data for chromosomes 2, 10, and 11 indicate that there are regions of frequent cross-over. One of these regions of frequent breaks is in the o region of chromosome 2.

Chiscon, J. A., and A. B. Burdick
Pleiotropism in three chlorophyll
mutants.

A previous research note from
this laboratory (TGC 11) stated
that our chlorophyll mutants,
when examined for chlorophyll

content, fall into 3 groups with respect to chlorophyll ratio. The largest group retains the same proportion of chlorophyll a to b as the wild types. A second group contains proportionally less chlorophyll a than the wild types. The third group contains proportionally less chlorophyll b than the wild types. This third group consists of only the yg₂, yg₅, and yg₆ mutants.

These same 3 mutants were found to have a greater reduction from wild type of total chlorophyll content than any of the other mutants which were examined. We have recently found that the quantity of free glutamic acid found in leaf material from these mutants is considerably higher than in the wild types. A few other mutants were found to accumulate glutamic acid, but these increases were slight and did not really approach the degree of accumulation evidenced in the three yg mutants.

When two of the three mutant factors (yg₂ and yg₆) were placed homozygously together in the same genotype, the resulting effect on glutamic acid accumulation appeared to parallel the effect of the same genotype on the chlorophyll ratio. Glutamic acid accumulation became less than in either single mutant and approached wild type. The chlorophyll ratio deviated from wild type less than in either single mutant and also approached the wild type.

There is little likelihood that these three mutants are in any way allelic. The three possible heterozygotes among them are all wild-type, and they (at least yg₂ and yg₆) appear to recombine freely. It is possible, however, that yg₂, yg₅, and yg₆ may be duplicate pleiotropic genetic units, whose manifold effects upon the tomato include at least the following:

- a. a severe reduction in total chlorophyll content
- b. an alteration of the normal ratio of chlorophylls a and b, with chlorophyll b becoming proportionally reduced
- c. an increased accumulation of free glutamic acid

The percentage of increase (I) or decrease (D) from wild type of optical density readings for glutamic acid.¹

LINE 018 MUTANTS

af - I - 5.7	li ₃ - D - 5.7	pg ₃ - I - 8.6
li - D - 5.7	md ³ - D - 14.3	yg ₅ - I - 40.0
li ₂ - I - 11.4	pg ₂ - D - 20.0	yg ₆ - I - 17.1

KOKOMO MUTANTS

pg - D - 50.0	yg ₃ - - - 0.0	(328) ² - D - 41.2
sy - - - 0.0	yg ₄ - D - 35.3	(338) ² - D - 44.1

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nd - D - 34.5	yg ₂ - I - 65.5
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1. obtained by paper chromatographic separation of free amino acids present in tomato leaf material
2. no gene symbol as yet assigned

Chmielewski, T. M., and C. M. Rick
Lycopersicon minutum.

Several accessions have been received of a new entity among wild tomatoes, which we

provisionally label L. minutum. This name was selected because this form is diminutive in most respects. All collections conform to the following description: Stems slender, copiously branched. Internodes rather elongate. In keeping with other tomato species, the foliage has a distinctive odor, in this instance suggesting burnt cheese. Leaves interrupted pinnate, but much less divided than those of most spp., in the size range of L. peruvianum var. humifusum, pseudostipules lacking. Leaf segments long petiolate, their margins serrate or undulate. Racemes simple without basal bracts. Pedicels often subtended by bracts, especially the basal one. Flowers small and inconspicuous, corolla 12 mm. or less broad. Style scarcely exerted. Fruit spherical, about 8 mm. diameter, minutely puberulent, whitish green at maturity with two darker green, sometimes purplish, radial stripes. Seeds small, about 1.5 - 2.0 mm. x 1.0 mm., broadly oblanceolate, hairy. All parts puberulent and glandular, but without large trichomes. The plants are highly self-fertile and set fruit abundantly in the greenhouse. In the field, and to some extent in the greenhouse, the plants are subject to severe wilting and collapse as if highly susceptible to vascular wilt diseases.

L. minutum is known from few localities: San Rafael and Chavinillo (Huánuco), Abancay (Apurimac), PERU. That these sites lie on the eastern slopes of the Andes is of special interest, for all other green-fruited tomato species occur west of the continental divide.

In two separate series of tests conducted in the greenhouse in different seasons of the year, crosses were attempted between L. minutum and representative forms of all other available tomato species and closely related species of nightshade. The only successful combinations were the following: L. minutum x L. esculentum and reciprocal; L. esculentum var. minor, L. pimpinellifolium, and L. hirsutum f. glabratum, all x L. minutum but not reciprocals; and L. minutum x S. pennellii, but not reciprocal. Seed yields were nearly normal in the first listed cross, but considerably reduced in the other combinations. Crosses with L. peruvianum, chilense, and hirsutum typ. failed, although combinations with L. minutum as female parent often yielded fully developed fruits with aborted ovules.

Hybrids with L. esculentum, L. esculentum var. minor, and L. pimpinellifolium were normal, vigorous, and fully fertile. Meiosis examined in the esculentum-minutum hybrid was normal. Hybrids with L. hirsutum f. glabratum were subject to withering of the stems as also manifest in hybrids between this species and L. esculentum. Meiosis was normal but fertility reduced. Hybrids with S. pennellii were normal, vigorous, but with fertility reduced to the same extent as in the esculentum-pennellii combination.

Segregations were tested in F_2 for a_1 , hl , c , d_1 , l_1 , and W_o received from the esculentum parent. In all cases recessive phenotypes were deficient, often significantly so. A recombination BC test was made with the a_1 - hl markers of chromosome 11. The individual segregation of the two genes was normal, but recombination in the hybrid (6.33%) was considerably lower than that of the control (11.04%).

The acquisition of L. minutum is of interest in several respects. As a source of germplasm for genetic and breeding studies it should prove a useful addition, for it clearly belongs with members of the esculentum complex and gene transfers to L. esculentum should not prove difficult. Taxonomically it is anomalous. According to the monographs by Luckwill and Muller it should belong with species of Eriopersicon, yet its genetic affinities are clearly

with Eulycopersicon. The need for a revision of the subgenera is therefore evident. The status of L. minutum itself is problematic. Like L. pimpinellifolium, it behaves genetically like a variety of L. esculentum, yet all three are separated from each other by large morphological hiatuses.

Clayberg, C. D. Inheritance and linkage of fruit stripe, Fs.

One characteristic of most or all species of Lycopersicon subgenus Eriopersicon is a dark

green striping originating at the point of stylar attachment on the ripe fruit and extending a variable distance towards the stem end of the fruit. The number of stripes on a fruit equals the number of locules and they are opposite each other in arrangement. The pigment of the stripe is located in the epidermis. Upon backcrossing this trait from L. hirsutum f. glabratum (C. M. Rick, Baños strain) into L. esculentum, we found that its inheritance is determined by a single dominant gene. In a predominantly L. esculentum background, fruits, in addition to having the stripes at the mature green stage of development, are highly pigmented, similar to hp fruits. When the fruit ripens, the stripes disappear completely or leave short yellow lines at the stylar end. It has not been possible to distinguish phenotypically Fs/Fs from Fs/+ plants. A marked and irregular deficiency of Fs plants has occurred in some backcross and selfed progenies, and an attempt to determine the cause of this is underway. However, lines have been obtained which are segregating in a regular fashion, and linkage data for these are presented here.

Family No.	Type	Marker "m"					Adjusted contingency	CO
			++	+m	Fs +	Fs m	Chi-square	
1	BC-C	u	0	28	31	0	54.97**	0
2	BC-C	u	2	33	23	0	46.65**	3 + 2.2
3	BC-C	u	0	24	36	0	55.91**	0
4	F ₂ -C	u	0	20	36	0	51.17**	0
5	F ₂ -C	u	1	12	42	3	33.75**	7 + 3.5
1	BC-C	t	5	23	20	11	11.40**	
2	BC-C	t	7	28	15	8	10.30**	
3	BC-C	t	8	16	30	6	13.42**	
4	F ₂ -C	t	12	8	31	5	3.68*	
5	F ₂ -C	t	7	6	31	14	0.439	

All five families confirm a tight linkage of u and Fs. The cross-over values were calculated by the maximum likelihood method, which also gives a combined best estimate of linkage value for u - Fs of $2 + 1.2$. The heterogeneity χ^2 for this estimate equals 39.27 and is significant ($\chi^2_{.05} = 9.49$, d.f. = 4), indicating that the five families are not segregating in the same ratio. Nevertheless, the estimate provides the best value for predicting future linkage results. Although u and t are both on chromosome 10, the progenies were too small to reveal gene sequence. The five families, which came from three sibling BC₄ plants in the previous generation, were also all segregating for a₁, e, mc, 2^{wf}, and y. None of these five markers provided significant contingency χ^2 values with Fs except for family 5, where a₁ gave an adjusted $\chi^2 = 4.60$. To conserve space none of the other values are presented here, although they are available to interested parties.

The striped fruit phenotype in L. hirsutum f. typicum (PI 127826) appears to be determined also by Fs or an allele of it. The following BC₃ family has been grown.

Progeny	Fs++	+ut	Fs u t	+++	Fs + t	+u+	Fs u +	+++
BC ₃ -C	54	46	0	4	12	23	0	0

The adjusted contingency χ^2 for Fs-u is 120.40 and for Fs-t is 27.05. The linkage value for Fs-u is $3 + 1.5$ which compares favorably with the combined best estimate previously obtained. These data also indicate a gene sequence of Fs - u - t.

Dempsey, W. H., and J. E. Boynton

Effect of time of day on
controlled pollinations.

A preliminary test to investigate effects of various environmental factors on pollination yielded such striking results that they

are considered worthy of presentation here.

On August 2, 1961, isolated Pearson (ms₂) and San Marzano (ms₇) plants supplied by C. M. Rick were hand pollinated every three hours for 24 hours in the field at Davis. Sixteen flowers for each of three crosses (P x P, SM x SM, P x SM) were pollinated with pollen that was collected the morning of the test and stored under favorable conditions in gelatin capsules. Pollen germination and rate of pollen tube growth were measured in all treatments by collecting 2 styles from each of the crosses at 6, 9, and 12 hours after pollination and observing pollen tube fluorescence by Martin's technique (TGC 9:38). Ten pollinations thus remained in each treatment, and at fruit maturity determinations were made for fruit number, weight, seed weight, and ripeness. Temperatures in the field on the day of treatment varied from a high of 91° at 5 PM to a low of 50°F at 7 AM. This range of temperatures is a few degrees below that of an average day at that time of year.

Both pollen tube growth and pollen germination were greatly reduced during the midnight-to-6 AM period. While pollen tubes were able to grow to the base of the style in 6 hours during the day and evening, 12 hours were required during the early morning hours. In the P x SM cross, two to three times as many pollen grains germinated as compared with the P x P or SM x SM pollinations. Also, the rate of pollen tube growth, particularly during the critical early morning period, was much more rapid than in the case of the hybrid combination.

Time	Temp. at 5 ft. level °F	P x P			P x SM		
		Number of ft. per 10 poll'ns	Mean ft. wt. gms.	Mean wt. seeds per ft. gms.	Number of ft. per 10 poll'ns	Mean ft. wt. gms.	Mean wt. seeds per ft. gms.
12 N	73	9	123	0.15	8	239	0.62
3	86	10	127	0.13	7	257	0.58
6	91	6	148	0.19	9	206	0.52
9	74	5	64	0.07	10	138	0.27
12 M	60	2	95	0.10	5	200	0.39
3 PM	54	2	85	0.12	6	135	0.24
6	52	5	94	0.08	7	111	0.15
9	60	1	3	0	8	190	0.37

The pollen tube results were borne out by the fruit-set and seed-set information presented in the above table.

In the P x P data the strong effect of environmental conditions at night, causing the reduction in pollen germination and tube growth, is reflected in reduced fruit and seed set. To some extent, this reduction was overcome in the hybrid cross, but the decrease at night is still clearly discernible. The SM x SM data are similar to those for P x P.

An interesting side observation to the experiment was the striking difference in maturity noted among the fruits. The fruits of the hybrid combination were notably earlier in maturity as measured by color. This metaxenia was possibly due to the greater number of seeds which these fruits contained and consequently increased metabolic activity of the fruit tissues. Also noteworthy is the strong correlation between number of seeds per fruit and size of fruit, the latter evidently being a function of the former.


Frankel, Rafael Tomato
inheritance punch card.


A simple hand sorted selection punch card was developed (see inserted sample specimen). It


was found, that the manual punch card system overcomes the inconvenience of searching for information recorded in tabulations and field books when the number of lines in a breeding program passes the 200 mark. It is suggested that TGC members adopt a standard punch card for tomato breeding material, which might accompany material exchanged between members. The purpose of this note is to arouse interest in this idea, and gather suggestions for an improved design of such a standard punch card.

In the card used by the author, recorded information can be selected for by manual separation of any combination of cross reference of the following:

1. Varieties, lines, or hybrids - by direct code.
2. Years grown - with years designation (13 years) - by direct code.
3. Fruit rating: Shipping quality - 4 grades.
Processing quality - 4 grades with purpose designation.
Organoleptic rating - 4 grades with type designation.
- all by direct code.
4. Fruit shape - 6 shapes - by numerical code.
5. Days to maturity (in 10-day graduations) - by direct code.
6. Days to peak yield (in 20-day graduations) - by direct code.
7. Locule number (2,4,6,7 locules) - by direct code.
8. Plant height in cm. (5 graduations) - by direct code.
9. Disease resistance rating in 7 degrees for 10 diseases - by numerical code.
10. Average fruit weight (in 25 gr. graduations) (back of card) - by direct code.
11. First letter of variety name, line or hybrid designation (back of card) - by alphabetical code.
12. 8 reserve holes for mutually exclusive information in a numerical code from 1-99, or any other arrangement of a key.
13. Genotype of 18 seedling and growth characters, 16 flower and fruit characters, and 6 disease resistance genes - by double-row coding as follows:

Heterozygous = shallow punch = 

Homozygous recessive = intermediate punch = 

Homozygous dominant = deep punch = 

In most selection categories the direct code system was employed in the card in order to facilitate minimum step extraction of cards according to a certain cross reference, and to avoid mutually exclusive classifications.

The card provides space for written information on:

- Fruit characters - 27 items
- Plant characters - 12 items
- Flower characters - 8 items
- General characters - 5 items

and for yield data, remarks, and a serial card number.

Griffing, J. B., and J. B. Langridge

Modification of recombination
frequency.

Much evidence has accumulated to show that recombination frequency is modifiable by both environmental and genetic

factors. The objective of this study, then, was to examine the influence of as many different factors as possible, in the hope of finding some method of increasing recombination frequency. Such a method would aid the plant breeder in overcoming the linkage barrier to the selection of desirable genotypes. Although we found no reliable chemical method of increasing recombination frequency, the methods and substances tested are here summarized in the hope that someone may pursue the subject further.

Since a large scale attack was contemplated, a technique permitting the rapid accumulation of data was necessary. Our approach was as follows:

Two tomato seedling mutants approximately 20 recombination units apart in chromosome 11 were used. These mutants, a₁ = anthocyanless and hl = hairless, can be scored on the hypocotyl a few days after germination. Seeds from plants doubly heterozygous in the coupling phase were placed in petri dishes for germination. The sprinkling of activated charcoal over the seed promoted rapid and almost complete germination. The germinating seeds were then placed in continuous light for the full development of anthocyanin. Scoring by aid of a low power binocular microscope was possible a few days after germination, or, in other words, in a week or so following fruit harvest. The above procedure was extremely efficient in providing a fast method of accumulating a vast amount of data.

1. Distribution of recombination frequency with regard to position of fruit in the plant structure.

The first objective was to examine the pattern of recombination frequencies as it occurs over the developing plant. In both pruned and unpruned plants, recombination frequencies were determined for individual fruits identified as to their origin in terms of clusters and branches. In the unpruned condition, 16,725 observations were taken. Significant differences were found among branches but the pattern of differences was complicated. In plants pruned to yield clusters only on the main stem, a significant negative correlation (based on approximately 10,000 observations) was found for recombination values and cluster number. This decline in recombination frequency, although significant, was not great. All plants in this study were maintained on a high plane of nutrition.

2. The effect of inorganic compounds on recombination.

Test plants were variously treated with EDTA, with foliar sprays of calcium and magnesium, and with nutrient solutions containing different levels of calcium and magnesium. Although over 35,000 seedlings were scored, none of these treatments could be shown to give unequivocal changes in recombination frequency.

In one experiment, the leaves adjacent to the cluster sampled were analyzed for calcium, magnesium, potassium, and sodium. A negative association between sodium and recombination frequency was the only significant result obtained.

The fact that poor nutrition may affect the frequency of recombination was indicated by periodic examinations of a test plant which was grown for over a year. In the absence of additional nutrients, the recombination frequency dropped from over 17 percent to 6 percent in six months. Mineral nutrients were then applied to the pot in which the plant was growing and the recombination frequency soon returned to 12 percent. However, in a further test involving 7,500 observations, foliar spraying with complete mineral solutions did not cause the treated plants to differ significantly from the untreated controls.

3. Effect of chemicals on recombination.

The effects on recombination of 21 chemicals when sprayed on test plants were assessed in a study involving over 27,000 observations. These chemicals (mutagens, hormones, anti-metabolites, metabolic inhibitors, etc.) were chosen for their known pronounced biological activities. A slight apparent increase was caused by IAA and benzthiazole oxyacetic acid. Barbituric acid, streptomycin and ribosenucleic acid lowered recombination frequency.

4. Effect of grafting on recombination.

In an attempt to determine the influence of the root system on recombination, a series of grafting experiments was made which yielded a total of 13,000 observations. The test plants were grafted onto another variety ('Sutton's Best of All'), onto Lycopersicon species (L. pimpinellifolium, L. peruvianum and L. hirsutum), and onto related genera (Capsicum annum, Solanum tuberosum and Datura stramonium). Grafted and ungrafted controls were included. Grafting of Solanum tuberosum produced the only significant result, which was a pronounced decrease in recombination frequency.

5. Genetic modification of recombination frequency.

The influence of the genetic background on the frequency of recombination between the two test loci was investigated by a series of backcrosses to eight different stocks which included L. pimpinellifolium, several cherry size varieties and several commercial types. After three backcrosses, the frequencies were examined in a study involving 19,671 observations.

It was shown that the different genetic backgrounds produced highly significant modifications in recombination frequencies - from 9.7 percent in L. pimpinellifolium to 21.0 percent in the 'Ponderosa' variety.

6. Summary.

Although this survey involved only a superficial study of an array of potential recombination stimulants, several points emerge:

- (1) The frequency of recombination is apparently modifiable by environmental factors (internal agencies producing ageing effects associated with poor nutrition) and also by genetic factors.
- (2) The possibility of increasing recombination frequency by the manipulation of environmental stimuli still remains unsolved.
- (3) Tomato breeders should maintain crossing material in good nutrition and if possible use flowers from the first cluster.
- (4) It might be wise to screen tomato breeding stocks for recombination frequency, and, in the long run, it may be wise to select for increased frequency of recombination.

Hagemann, R. Instability
at the yv locus.

By means of X-irradiation a
green-yellow variegated plant
originated from the tomato

variety Condine Red. The mutant line, to which this plant gave rise, consists of seedlings with pure green cotyledons, seedlings with green-yellow variegated cotyledons, and pure yellow seedlings. The variegated seedlings show all gradations of variegation, so that the pure green and the pure yellow seedlings are really the extremes.

The yellow seedlings gradually develop into yellowish-green plants; in the greenhouse they even become light green. They form flowers, but they do not produce seeds after selfing, because they are completely male-sterile.

The green and the variegated seedlings develop into variegated plants with yellow spots or larger sectors on a green background. All gradations of variegation are also found in the foliage leaves. Variegated plants sometimes form pure yellow branches, which are male-sterile just as the yellow plants.

The progenies of variegated plants (both from green and from variegated seedlings) segregate into green, variegated, and yellow seedlings (the sum in 1961: 556 green, 4503 variegated, 1960 yellow seedlings). In the progenies of plants, which had variegated cotyledons, there are, in general, many more yellow (46%) and less green (6%) seedlings than in the progenies of variegated plants which had green cotyledons (24% yellow and 25% green).

The F_1 plants of the crosses

- (1) yellow plant x Condine Red,
 - (2) yellow branch of a variegated plant x Condine Red,
 - (3) variegated plant x Condine Red and reciprocal,
- are pure green.

All F_2 generations of (1), (2), and some of (3) segregate into green and yellow plants (no variegated!). The greater part of the F_2 generations of (3) segregate into green, variegated and yellow seedlings. The proportion between green and yellow resp. between green and (variegated + yellow) corresponds to a monohybrid ratio, but with a deficit of the mutant types, especially of the yellow ones.

Crosses between yellow plants of our mutant line (y) and the marker stock yv, dl, al, wt (kindly supplied by Watkin Williams) revealed that the yellow plants are homozygous for an allele of yv (Robinson and Rick, 1954), which has been localized in chromosome 6 (IV) (TGC 10:5, 1960). The hybrid is yellow virescent in phenotype and self-fertile.

The following formal explanation is given for the results:

The yellow plants, as well as the yellow branches of variegated plants, are homozygous for the allele yellow virescent^{masculosterilis} (yv^{ms}). The cross yv^{ms} yv^{ms} x + + (= Condine Red) gives green F_1 plants (+ yv^{ms}), and in F_2 there is a segregation into green (+ + and + yv^{ms}) and yellow plants (yv^{ms} yv^{ms}) [Cross (1) and (2)].

The variegated plants contain the allele yellow virescent^{mutabilis} (yv^{mut}), which determines green plant color just as +, but mutates in vegetative tissues regularly to yv^{ms}.

Almost all seedlings with variegated cotyledons have developed from yv^{mut} yv^{ms} zygotes. In such cells the mutation of yv^{mut} produces a yv^{ms} yv^{ms}

(yellow) constitution. Whether this mutation causes a yellow spot on a leaf, a larger yellow sector or a pure yellow branch depends upon the developmental stage in which the mutation takes place. Almost all seedlings with pure green cotyledons have developed from $yv^{mut}yv^{mut}$ zygotes. Thus two mutational events (in both homologous loci in a cell or its descendants are necessary to produce a yellow spot. This usually does not happen before the leaves are formed. These differences between the two types of variegated plants explain why the first type produces more yellow and less green seedlings in its progeny than the second (as mentioned above).

The cross (3) variegated plant [$(yv^{mut}yv^{mut} + yv^{mut}yv^{ms} + yv^{ms}yv^{ms})$] x Condone Red [+ +] gives two types of green F_1 plants. The greater part of the F_2 generations (progenies of + yv^{mut} plants) segregates into green, varietaged and yellow seedlings. Some F_2 generations (progenies of + yv^{ms} plants) segregate only into green and yellow seedlings.

Within the multiple series at the yv locus there are the following relations of dominance:

$$+ > yv^{mut} > yv > yv^{ms}.$$

The event which underlies the hereditary change, formally called mutation of yv^{mut} to yv^{ms} , may be a

- (a) gene mutation,
- (b) loss of a part of chromosome 6 (induction of a deficiency),
- (c) loss of a fragment, which together with a deficient chromosome 6 gives a normal phenotype (as described in the "flaked" mutant by Lesley and Lesley in Genetics 46:831, 1961). Cytological studies are under way to find out which of these possibilities is realized in the present case.

Hansen, D., C. M. Rick, and
J. E. Boynton Linkage tests
with mutants of Stubbe group
III.

Mutants of this group are being
systematically screened for
linkages following procedures
outlined in previous notes
(TGC 10:38; TGC 11:22).

Mutants with dependable seedling expression are selected for this study. To date linkages have been encountered for 6 of these mutants. Results of all completed tests between these 6 and a series of testers are summarized with similar results with two mutants of the Stubbe I series in the note by Rick and Martin (p. 43). Segregations for the linked combinations are presented in the following table. Crossover values are given only for the most dependable segregations.

The data permit positioning only for one (spa) of the six mutants. Limited as the data for spa are, its locus must be between dl and l_1 . The indicated distances fit surprisingly well with the existing map, but a position for spa midway between the two markers can only be tentative. In the $spa-l_1$ segregation some doubt existed about the expression of l_1 in the two $spa-l_1$ segregants; it is possible that no recombinants appeared and that the map distance between the two genes might be considerably less than estimated. Obviously much more needs to be done here, and spa will be entered in multipoint tests. Stocks of these mutants and pertinent segregating progenies have been transmitted to cooperators in charge of the respective chromosomes.

Combination	++	+t	m+	mt	Adjusted contingency Chi-square	co.	Mean co.
bip-d ₁	293 178 211	114 62 98	119 62 78	27 5 1	4.63 9.31 29.11	11.5	
gri-e	299	119	130	20	12.87	37.0	
id-a ₁	214 242 194	103 104 104	97 71 97	15 3 15	14.20 20.36 17.25	34.5 21.0 33.0	29.5
mu-c	274	108	78	0	25.47	0-8	
oli-H	179 229	81 84	80 66	8 1	15.67 18.98	14.0	
spa-l ₁	182	102	58	2?	23.41	17.0	
spa-dl	157	72	40	1	13.39	16.0	

Joubert, T. G. la G. A new jointless gene, j_2^{in} .

This new jointless gene (j_2^{in}) was first reported in TGC 11:13. It differs from j_1 and j_2 in that

it shows a normal pedicel joint, but the abscission layer is absent in the mature stage. This jointless condition is delayed until the fruits reach a fair stage of enlargement. It is associated with leafiness, but in most of the cases examined this leafiness is also delayed and only found on the third or fourth truss on the plant. When crossed with normal jointed varieties j_2^{in} behaves as a monogenic recessive. It behaves as a dominant when crossed with j_1 and j_2 .

The results of a few crosses are given below:

No.	Cross	F ₂ segregation			
		+	j_2^{in}	j_1	j_2
1	$j_1 \times j_2^{in}$	13	58	22	--
2	$j_2^{in} \times j_1$	28	95	30	--
3	$j_2 \times j_2^{in}$	2	59	--	22
4	$j_2 \times j_2^{in}$	0	94	--	23
5	$+ \times j_2^{in}$	77	25	--	--

These data show recovery of + phenotypes in the F₂ of $j_1 \times j_2^{in}$; whereas, with few exceptions, none are recovered from the F₂ of $j_2 \times j_2^{in}$. The new mutant thus appears to be a new and different allele of j_2 and is assigned the symbol j_2^{in} for jointless-2 with incomplete action.

If the pedicel joint of j_2^{in} plants are examined in F₂, a variation in the degree of woodiness of the abscission layer is quite obvious. Plants with slightly, moderately and very woody abscission layers are present. Selections of these different types in F₂ segregated in F₃ as follows:

Cross	Character selected in F ₂	F ₃ segregation		
		+	<u>j</u> ₂ ⁱⁿ	<u>j</u> ₁
<u>j</u> ₂ ⁱⁿ x <u>j</u> ₁	Slightly woody	21	35	4
<u>j</u> ₂ ⁱⁿ x <u>j</u> ₁	Moderately woody	4	78	0
<u>j</u> ₂ ⁱⁿ x <u>j</u> ₁	Very woody	1	73	1

Kerr, E. A. Another virescent gene. What shall we call it?

A virescent mutant arose in 1957 as a single plant in a breeding line derived from Fireball x

(Oxheart-Pritchard-L3700II-Red Jacket). Under winter greenhouse conditions the new leaves are pale but frequently the mutants are indistinguishable from normal under summer conditions in the field. Growth and fertility are normal. F₂ populations give good 3:1 ratios. Preliminary tests indicate that there is no close linkage with a, ag, c, d, e, l₁, mc, r, sp, t, tf, u, wf, y, or yv. Further tests are being made with genes on chromosome 2 as there was a slight suggestion of linkage with d.

In preliminary tests to establish that this is a new mutant, it has been designated as vir I5, i.e., a virescent mutant from the breeding line I5. However, vir is not an acceptable symbol for this gene as it has previously been used for viridis--a dark green mutant with third-order leaf divisions. The symbol v was used for a virescent mutant in 1933. This mutant apparently was semi-lethal and has been lost. The present mutant is definitely not a mimic of v. In many respects it resembles op and t^v. It may resemble dis, luc, va, or ver but these have not been compared.

The system of gene symbolism is becoming too complex. Many genes that are mimics or have similar phenotypes have been given dissimilar designations. The following genes all give green-stemmed plants: a, ae, af, ag, ah, ai, al, ao, and aw. The following have third-order leaf divisions: vir, tp, ff and a new mutant which we are now testing. A host of mutants have "light-green", "yellow-green", "gray-green", and "pale-green" foliage.

The work of establishing the linkage relationships in specific chromosomes has been assigned to specific people. I propose that, as soon as possible, we do something similar in naming new mutants and in re-naming many of the old ones. To this end a committee--possibly the committee on nomenclature--could assign one or two people to compare all present and future mutants of, for example, one of the following: stem growth, leaf color, leaf shape, flower characters, fruitfulness, mature fruit characters. These people would then be in a position to clarify the designation of mutants and could either assign names and symbols for mutants or at least make recommendations for their naming.

The symbol v₂ has been tentatively assigned to this mutant (Ed.).

Kerr, E. A. Light green-5, a second marker for chromosome 5.

This mutant was first noticed in the spring greenhouse crop of 1959 in a line derived from

PI#126954 (L. pimpinellifolium) x (Vetomold-L. hirsutum) outcrossed four times to Tuckqueen. It has normal fertility and almost normal growth. As it closely resembles lg₁ on chromosome 10, the gene symbol lg₅ is proposed. One

F₂ population with the marker c d gs h j₁ r gave 250+ : 63 lg₅, i.e., a slight deficiency of lg₅ plants. No linkage was indicated with c d h j₁ r but a crossover value of 12 was obtained with gs (+166 : +gs 84 : lg₅ + 62 : lg₅ gs 1). This indicates that we now have a second gene on chromosome 5. As it can be easily classified shortly after the plants are set in the field, it should prove useful as a marker for this "neglected" chromosome.

The F₁ of lg₁ x lg₅ is normal and 4 small F₂ populations gave 81+ : 52 lg. No extremely pale plants were noted. This suggests that the homozygote lg₁ lg₅ is lethal or that there is reduced viability.

A small repulsion F₂ with marker LA159 (a e mc t u wf y) gave unexpected results. It produced 44+ : 11 lg₅ : 4 yellow-green. The yellow-green plants had normal growth and fertility and proved to be homozygous recessive. Preliminary F₂ tests indicate that this yellow-green phenotype is the homozygous double recessive of lg₅ and a hypostatic gene from LA159.

Lesley, J. W. Recombination data with blue-green (be).

Blue-green described in Genetics 41:575 (1956) and in Am. Jour. Bot. 45:598 (1958) is associated

with a short inversion in a chromosome not yet identified. Blue-green is not easily recognized in the greenhouse at temperatures ranging from 65° to 85°F but at Riverside, California, is easily identified by plant size and color after a few weeks growth in the field. It tends to wilt more than normal sibs but is quite fruitful. Blue-green is less easily identified in combination with genes affecting plant size and green color so that such data are less reliable. The following F₂ data suggest independent recombination but may help in future linkage studies. Seed of blue-green is available from TGC sources. Ratio +:be was $\frac{1}{3.3}$.

Combination	Phase	++	++	m+	mt	χ^2_L	Ref.
m	t						
be	j	C	96	28	42	10	0.4
		R	60	27	9	13	0.5
be	sn	R	76	34	31	12	0.1 TGC 11:15, 1961
be	d	R	149	40	48	28	7.9 contrary to phase
be	c	C	36	9	11	7	2.7
		R	102	33	26	9	0.02
be	sp	R	18	4	6	4	2.0
be	r	R	70	15	14	5	0.8
be	l	R	65	21	19	6	0.0
be	a	R	68	21	8	14	11.5 contrary to phase
be	y	R	70	25	20	3	0.1
be	al	R	39	8	6	5	3.4 contrary to phase

Lesley, Margaret M. Change of fragment size in "flaked" variegation (submitted by J. W. Lesley).

It was reported (J. W. and M. Lesley, Genetics 46:831-844, 1961) that "Flaked" plants have either one or two minute extra chromosome fragments. A lightly

flaked plant with one, and a moderately flaked plant with two, much larger extra fragments have been found in plants derived from selfing flaked. When these two extra fragments are present at diakinesis or M_1 , they show a definite attraction for one another, but close pairing is rare. As a consequence their orientation on the spindle is erratic, and occasional loss of one or both fragments should occur. At pachytene two short extra rods are easily demonstrated but they are rarely closely associated. When only one such fragment is present it shows a strong tendency to associate with a pair of chromosomes, possibly number eleven. A dimorphic pair may result in which the centromere of one member is increased in length and has a large pycnotic body at its center. This pycnotic body is nearly the size of the free fragment. It may also be attached to a whole chromosome (No. 11?) by one slender strand. A drawing of this chromosome at pachytene measures 5 inches in length, the free fragment only measures three sixteenths of an inch. In one cell at pachytene a clear ring was found in addition to 12 pairs. This ring might have originated from two breaks near the centromere, one on either side of the central pycnotic body in the heteromorphic pair. Both rings and telocentric rods-fragments have been described as giving rise to larger or smaller fragments.

Lyall, L. H. Necrotic (ne) phenotypes in crosses with the Atom variety.

Atom tomato is a very early type with a small determinate plant and has the ability to flower and set its small fruit under cold temperatures.

At Ottawa in 1961 F_2 progenies from 3 crosses, each with the variety Atom as one parent, segregated for ne in the proportion of 13 normal : 3 necrotic plants as follows:

Cross	Normal	Necrotic	P. for 13:1 ratio
0.60M x Atom	120	30	0.7
0.60R x Atom	99	21	0.5 to 0.7
0.60D x Atom	126	23	0.3

The expression of ne requires the presence Cf_2 (C. fulvum resistance) and necrotic plants should be $Cf_2/ne/ne$. The gene ne is carried by most varieties of L. esculentum and it would appear that Cf_2 must be carried by the Atom variety.

Atom was originally obtained by us from Thomas Cullen & Sons, Witham, Essex, England.

Mathan, D. S., and R. F. Stettler
Age of seed effect in homozygous lanceolate seedlings.

As previously reported (Science 131:36-37) La+ plants, upon selfing, segregate typically 1 normal: 2 lanceolate: 1

homozygous lanceolate, the latter class often being somewhat deficient. In the early stages after germination two distinct forms can be recognized among

the LaLa genotypes, i.e., "modified" and "reduced." Later on, some of the "modified" seedlings often give rise to "narrow" plants while others never develop beyond a short first leaf. "Reduced" plants, on the other hand, always remain a short cylindrical stub without a trace of cotyledons or plumule. (For further description of these phenotypes see the Science paper.) Earlier data had suggested that the proportion of "reduced" plants among LaLa genotypes was a function of age of seed. To further investigate this question several large seedlots were extracted from the same culture and then aged under normal storage conditions. At various intervals, samples were taken and either dissected or germinated in sterile petri dishes.

Except for ++ and La+, which could not be distinguished from each other, all phenotypes could be recognized clearly in dissected seed. Moreover, there was a close agreement between data from dissection and data from germination of seed drawn from the same lot, which indicated that the germination process did not induce any significant shift in the relative frequency of the types distinguished. Careful scrutiny under the dissecting scope revealed an additional form among LaLa seedlings characterized by incomplete reduction of the single cotyledon. Since these "almost reduced" seedlings invariably behaved like truly "reduced" ones in their subsequent development they were included in the latter group for means of comparison. Preliminary data are summarized in the table below:

Segregation of phenotypes in progenies of selfed La+ stored for various lengths of time.*

		Period of seed storage			
		32 days	119 days	154 days	190 days
++ and <u>La+</u>		158	447	786	680
<u>LaLa</u>	non-reduced	36	86	106	75
	reduced	10	70	139	121
-----		-----			
Total		204	603	1031	876
Reduced in % of all <u>LaLa</u>		22	45	57	62

*In each lot all seeds were analyzed; those which failed to germinate were dissected and classified.

The data clearly indicate an increase of the fraction of "reduced" seedlings with length of seed storage. Moreover, this conclusion is supported by similar observations in related but different cultures of the same line. Hence, during the process of aging some of the "modified" seedlings seem to be converted into "reduced" ones. This conversion is presumably the result of a process by which the single cotyledon of the "modified" is progressively absorbed. The various intermediate forms observed are assumed to represent different stages in the sequence of this process. More critical techniques are presently being devised to subject this phenomenon to a closer examination.

Mauli, Chandra, and N. K. Notani

A survey of naturally occurring seedling mutations in the tomato populations around Bombay.

Progenies from 45 tomatoes collected from the local market were screened for pollen sterility, chromosomal aberrations, aneuploidy and gene

mutations. Although a great deal of variation for pollen sterility was observed, no chromosomal aberrations or extra chromosomes could be detected. However, one line clearly segregated for a chlorophyll deficiency which seemed simply inherited. Assuming complete self-pollination and a gametic mutation rate of μ ($A \rightarrow a$), the probability that a homozygous diploid individual will become a heterozygote through the mutation of one of the two alleles is 2μ in any generation. Since the proportion of heterozygotes in a selfing population is halved in each generation, the proportion of heterozygotes due to mutation pressure in such a pure line is (Haldane, J. B. S., 1936, J. Genetics 32:375-391):

$$2\mu \left(1 + \frac{1}{2} + \left(\frac{1}{2} \right)^2 + \left(\frac{1}{2} \right)^3 + \dots \right) = 4\mu \dots (1)$$

If the mutant gene is lethal in homozygous condition, a heterozygote on selfing will produce $\frac{1}{4} AA + \frac{2}{4} Aa$. Therefore, the proportion of heterozygotes due to recurrent mutations in a selfing population is:

$$2\mu \left(1 + \frac{2}{3} + \left(\frac{2}{3} \right)^2 + \left(\frac{2}{3} \right)^3 + \dots \right) = 6\mu \dots (2)$$

Assuming even the highest rate of natural mutation known in plants, that for R locus in maize of 5×10^{-4} , the number of heterozygotes expected would be $4 \times 5 \times 10^{-4}$ or 1 in 500 plants, which is still about one-eleventh of what is observed here. It appeared, therefore, that this might be fortuitous.

A second collection was, therefore, made from Panvel, a place about 40 miles from Bombay. 151 progenies from single fruits were screened and two mutations, one for much reduced anthocyanin (segregation 21 wild: 7 reduced anthocyanin) and another one for hairlessness (23 wild: 8 hairless), segregated out. This gives an average gametic mutation rate of about 1 in 38 for the two viable mutations and 1 in 8 for the lethal mutation--an extremely high mutation rate (two populations considered different). It is possible that the apparently high mutation rate is due to often cross-pollination in tomato under these conditions.

Notani, N. K. Probable genetic location of the centromere on the linkage map of chromosome 8 in tomato.

Genetic location of the centromere on the linkage map of chromosome 8 in tomato was attempted. The method relied on the chance generation of a

secondary trisome from a primary one, resulting in the altered segregation of the gene markers borne on the two chromosomal arms. Stocks with three markers, dl (dialytic), bu (bushy) and l₁ (lutescent) were crossed to the primary trisome 8. The genetic map positions of the three markers are as follows: dl at 40, bu at 49 and l₁ at 76. From four F_2 progenies over 1600 seedlings were classified for the three markers. Results are relatively consistent from progeny to progeny and there is no evidence of any marked imbalance of the ratios which would indicate that a secondary trisome had been generated (Table 1). Frequency of dl seems to be very high as is somewhat true for other markers also, but it is extremely unlikely that this is indicative of any telocentric or an isochromosome in as much as all the four progenies seem consistent in this respect.

The classification, taken singly, of the markers is as follows: dl 23.73%, bu 15.22% and l₁ 17.51%. This relative segregation seems to be obtained consistently from progeny to progeny. Consequently, it seemed tempting to assume that the segregation of these markers (from a trisome) is proportional to their distance from the centromere. Advantage was taken of the fact that the known genetic data impose a constraint and specify the gene order with bu as the middle gene. Thus the centromere can take only four positions: 1) Sfa dl bu l₁, 2) dl Sfa bu l₁, 3) dl bu Sfa l₁, 4) dl bu l₁ Sfa (Sfa abbreviation for Spindle Fibre Attachment).

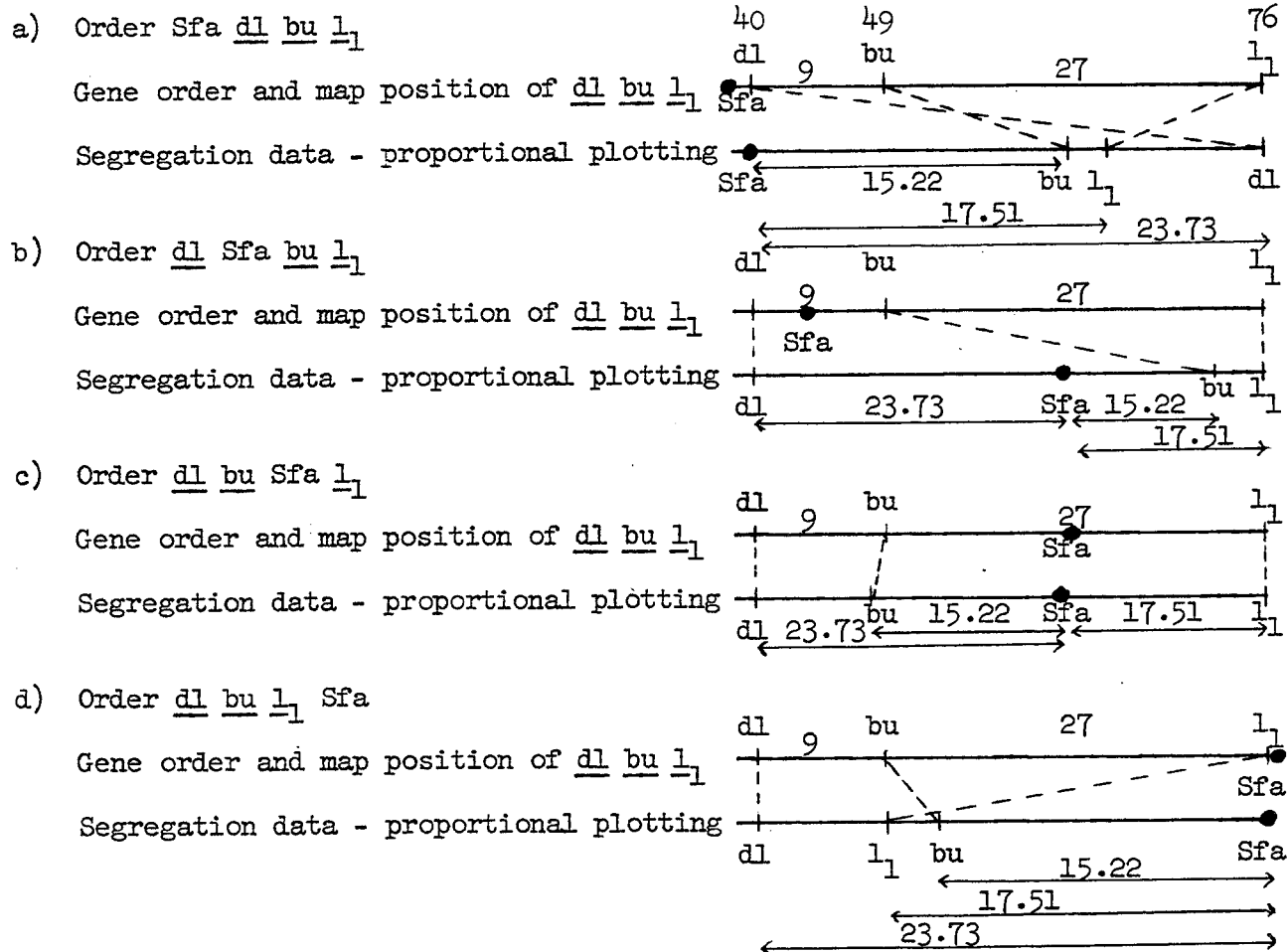
Thus it can be seen that the order Sfa dl bu l₁ is not possible because dl segregation should then be the lowest being nearest to the centromere. The second possible order dl Sfa bu l₁ is also not likely because the genetic distance between dl and bu is 9 map units, with segregations of 23.73% and 15.22% respectively, whereas l₁ with over 27 map units gives a segregation of only 17.51%. The third order dl bu Sfa l₁ seems to satisfy the genetic data and the segregation data. With 27 map units distributed over bu Sfa l₁ (segregation respectively 15.22% and 17.51%) and with dl 9 units distal to bu, it can be seen that both the segregation data and the proportionality data are satisfied. The fourth possible order - dl bu l₁ Sfa - is excluded by a similar argument that l₁ being nearer to the centromere should give a smaller proportion than bu which is contrary to what is observed here. Thus the position of the centromere seems to be between bu and l₁ with centromere slightly nearer bu than l₁ (See Fig. 1). It should, however, be emphasized that proportions of all three markers are far too numerous even when one considers maximum random chromatid segregation, the reason for which remains obscure. The data are, however, meaningful if the legitimate assumption is made that the double reduction is proportional to the distance of a gene from the centromere and that the total segregation is the sum of a proportionality component and a constant component. Also, these deductions do not consider any localized interference phenomena or changed dominance relationships.

Proof for this could come from an experiment in which a multiple linked recessive stock is crossed on to an 'asynaptic' plant (but having the wild type genotype for other markers). If the asynaptic generates an isochromosome or a telocentric, the F_1 plant should have the phenotype of some of the recessive markers, owing to the loss of the chromosomal arm that carried the dominant wild type markers. The F_1 screening can be done phenotypically and the presumable isochromosome or telocentric carrying plants can be confirmed cytologically.

Table 1. Segregations observed in the F_2 of the cross Triplo 8 x dl bu l₁

Plant No.	+++	dl ++	+bu +	++l ₁	dl bu +	+bu l ₁	dl+l ₁	dl bu l ₁	Total
3 - 1	337	71	27	36	14	39	13	9	546
3 - 2	87	26	9	14	5	7	2	3	153
3 - 3	253	73	17	38	5	14	7	1	408
3 - 4	239	119	49	41	7	31	19	10	515
Total	916	289	102	129	31	91	41	23	1622
% of Total		23.73	15.22	17.51					

Fig. 1.



(Grateful thanks are due Prof. C. M. Rick for his encouragement and for help given by way of materials and consultation in doing this problem.)

Prend, J. Effect of gibberellin
upon male sterile plants of
ms₂₃ Early Pak.

Since gibberellins are known to
induce flowering in a variety
of plants including staminate
flower formation in gynococious

cucumber, interest was focused here on the possible effect of this substance
on genetically male sterile plants.

In preliminary tests conducted during the summers of 1960 and 1961, an
application of potassium giberellate (Gibrel, a solution of potassium
giberellate, Merck) in concentration of 250 ppm made upon male sterile plants
of ms₂₃ Early Pak (kindly made available by Dr. C. M. Rick) caused considerable
elongation of the style and also appeared to have stimulated parthenocarpic
fruit formation.

The chemical was applied early in August and late in July respectively,
in 1960 and 1961 upon 3 and 5 male sterile plants at a rate sufficient to wet
the plants with no excess to drip off. An equivalent number of plants were
left untreated. In about 7 to 10 days following the application, style
elongation approaching nearly twice the length of those untreated was observed
on flowers of all treated plants. This condition prevailed for about 5 to 6
days and flowers developing after that period appeared to be normal.

Summary of Fruit and Seed Production

Year	No. of Parthenocarpic Fruit per Plant		No. of Fruit per Plant With One or More Seeds	
	Treated	Untreated	Treated	Untreated
1960	9.0	1.9	0.0	1.7
1961	15.4	1.4	6.0	6.6

The fruit set examined later in the season revealed that the number of parthenocarpic fruit on the treated plants was considerably higher than the number on the untreated. The recorded number of seed-bearing fruit (apparently the result of natural cross pollination) on the treated plants showed no increase over the untreated. While gibberellin increased the length of the style, it evidently did not restore self fertility.

Rick, C. M. Inheritance and linkage relations of compact, cpt.

Discovered as an unfruitful plant (2- 377) in the variety XL Pearson, this mutant has consistently shown the same

features in several seasons of field culture. Although few fruits were set on the source plant, they were moderately seedy, and all cultures planted from this source have bred true for the following phenotype. The plant differs strikingly from normal in its assuming a uniformly compact mound of growth. Unlike the typical straggly growth of a tomato plant, the periphery of cpt plants in midseason is smooth and well rounded. This compactness is vested in a slower rate of growth and marked increase in degree of branching. The main branches of the plant produce laterals from nearly every leaf axil; scores of shoots can be counted in the periphery of growth. In spite of its slower growth rate, cpt exceeds normal in total growth by virtue of its not being impeded by heavy fruit set.

Leaves of cpt are so different from normal in color and form that they alone are sufficiently diagnostic for accurate field identification. Leaf color deviates in a yellow-green direction. Leaves are smaller, have fewer small segments and, as a result of foreshortening of the main axis, the segments are brought into closer approximation, lending a unique appearance to the plant. Stems are more lax and leaves more limp, imparting a soft texture to the plant.

Flowers and fruits are modified little or none in cpt. Pollen is abundant and the cause of unfruitfulness is unknown. Plants set about 5% of a normal fruit load. Attempts to identify cpt in the seedling stage have been unsuccessful.

Outcrosses to normal have yielded only normal plants. Segregation in nine F_2 families from crosses to different lines shows normal monogenic recessive inheritance (Total of 9 families : 1212 + : 386 cpt; $\chi^2 = 0.57$, $p > 0.3$; Het. $\chi^2 = 13.54$, 8 df, $P > 0.05$). Linkage tests with 16 markers are summarized in the accompanying table. Chi-square tests were made only for data sets that show marked deviations in the direction of linkage.

Significant deviations are revealed for dl, l₁, H, and tf. Those for dl and l₁ are mutually confirmatory, while those for H and tf are on the border-line of significance. Moreover, the linkage with dl and l₁ was confirmed by trisomic segregation in triplo-8 (75 2N+:3 2Ncpt:24 tr-8+:1 tr-8 cpt), while

the trisomic segregations for chromosome 7 (47 2N+: 12 2Ncpt: 24 tr-7+: 7 tr-7cpt) and chromosome 10 (36 2N+: 13 2Ncpt: 6 tr-10+: 2 tr-10cpt) discounted the possibility of a linkage with either H or tf. A locus for cpt on chromosome 8 is therefore a certainty. The linkage data fit reasonably well with the existing map, yielding the following relationship: $\frac{dl}{40} - 11 - \frac{cpt}{51} - 26 - \frac{l_1}{77}$.

Tester		<u>+</u> <u>+</u>	<u>+</u> <u>t</u>	<u>cpt</u> <u>+</u>	<u>cpt</u> <u>t</u>	Adj. con. Chi-square	Co.
a ₁	F ₂ -R	160	63	53	24		
ah		169	62	61	22		
au		213	49	45	13		
c		153	70	56	21		
d ₁		95	42	14	8		
		161	62	63	14		
dl		153	77	83	1	32.648	11
e		182	61	55	20		
j ₁		184	59	60	15		
l ₁		157	66	72	5	15.655	26
H		168	75	62	13	4.588	
r		162	53	58	15		
rv		216	27	65	10		
tf		185	77	53	5	9.685	
wf		182	61	52	23		
y		170	45	54	19		
yv		184	46	73	11		

Rick, C. M. Inheritance and linkage relations of Hirtum, Hrt.

Hirtum is characterized by a striking increase in the density of large trichomes on stems and

peduncles. The hairiness is so dense in field-grown plants that the stems have a fuzzy appearance. Unlike Wo and its alleles, Hrt has leaf hairs that do not seem to be modified. Other manifestations of Hrt have not been detected. Attempts to distinguish it in the seedling stage have not been successful. Two sources are known for Hrt: (1) LA501 a single spontaneous rogue encountered in experimental plantings at Shafter, California; (2) LA393 a primitive cultivated tomato purchased in the market at Chiclayo, Peru. All of the 4 F₁ plants from a cross between these two lines and all of the 58 F₂'s showed the same hairy phenotype, thereby revealing an allelic relationship between the responsible genes of the two. Plants of F₁ Wo^m x Hrt from both sources showed intermediate phenotype for both genes and the F₂ showed independent segregation for both, 5/83 plants being of normal phenotype in respect to both characters.

Crosses between Hrt and + yield hybrids that are noticeably hairier than normal and F₂ segregation in 8 families provides a good fit to monogenic expectations² when hairy is classified against normal (692 Hrt: 220 +; $\chi^2 = 0.33$, $p > 0.5$; $\chi^2_{het} = 9.54$, 7 df, $p > 0.2$). Distinguishing between Hrt and Hrt/+

in these families has thus far not been successful, although gradations in the degree of excess hairiness are evident. Extensive linkage tests have not been attempted, but a cross intended to combine Hrt with La revealed the following interesting relationship:

	+/+	Hrt/+	Hrt/Hrt
+	3	5	13
La	22	36	2

Although these are small figures and the separation between Hrt/+ and Hrt/Hrt is debatable, the test for independence in a 2 x 3 table gives a chi-square of 30.83, which is very highly significant. This indication of linkage with La is currently being tested by a cross between Hrt and La-deb.

Rick, C. M. New male-sterile mutants.

Following past practice, we have been investigating new male-sterile mutants only when

they appear in new varieties or present some features of unusual interest. Of the four mutants reported here, ms₃₀ in San Marzano and ms₃₅ in A-1 are reported because they differ in interesting ways from mutants previously reported in those varieties, and ms₃₃ and ms₃₄ constitute the first male-sterile mutants of the variety VF-11³³ Pearson. The symbol ms₃₀, previously applied to a mutant of A-1, is reassigned to the new mutant of San Marzano for the following reasons. In the two seasons following its introduction, the original ms₃₀ proved undependable because it produced variable amounts of fertile pollen and selfed seeds. Unquestionably a hereditary defect exists in this line, but it clearly is not a male-sterile mutant. Unless some interest should develop in this mutant (2-443) it will not be reassigned a symbol but will be kept in the inactive list.

As in the past, it is assumed that the new mutants reflect different genes than those reported previously for the following reasons. The phenotype of ms₃₀ is completely different from any previously described mutant and therefore must represent a new mutation, most likely at a new locus. The same reasoning applies to ms₃₅. For the mutants in VF-11 there can be no dispute concerning their recent origin because they were discovered in this variety only four generations subsequent to the original single plant selection. Since they differ from each other in drastically different phenotypes, it is also likely that they represent mutants at new and different loci. Previous allele testing with the San Marzano series revealed all completely male-sterile mutants to be non-allelic.

It is interesting to note that two different mutants were found in the new variety VF-11 in 1959, when no more than four generations, and possibly only three, had intervened since the original single plant selection originating this variety. Since no sterile plants had appeared in the second generation it follows that the mutations giving rise to these new mutants must have occurred at some time during development of the second or third generations. A third sterile plant had been found at the same time with phenotype identical with that of ms₃₄, its F₂ segregating for the same phenotype. Although their identity has not been proved, it is very likely that both original plants had the same mutant gene and were segregants from the same original mutation. The chances for such repeated recovery would be much greater in such a recent variety, in which only a limited number of plants sired the examined population.

Descriptive information for these new mutants is presented in the following table. From the standpoint of ease of identification and exposure of stigma, all four mutants are satisfactory for hybridization. Within the limits of a single season's data, the seed yields indicate normal fertility for ms₃₃ and ms₃₅ and reduced fertility for the other two mutants, qualifying the former pair as better parents for large-scale crossing.

Mutant	Acc. no.	Variety	Flower size	Percent flowers with exposed stigmas	Stamens	
					Color	Size and Shape
ms ₃₀	2-455	San Marzano	Much reduced	100	greenish to pale yellow	highly modified; very slender, seldom connate; anther tube spreads basally.
ms ₃₃	2-511	VF-11	Normal but distorted	100	green or slightly yellow	highly modified and irregular; basally expanded but distally filamentous; some anthers connate and often adnate to style
ms ₃₄	2-513	VF-11	Normal	100	paler; comparable to corolla color	short, shrivelled, and partially dialytic basally
ms ₃₅	2-517	A-1	Smaller than normal	100	paler; comparable to corolla color	uniformly very much reduced; usually connate; slender symmetrical tube

Mutant	Time of breakdown	Meiosis	Contents of mature anther	Additional features
ms ₃₀	early prophase of meiosis	early stages appear abnormal	empty	very low female fertility
ms ₃₃	early prophase of meiosis	early stages appear abnormal	empty	distorted gynoecium
ms ₃₄	tetrad and early microspore	normal but possibly delayed	empty PMC; disintegrating tetrads; rare stainable pollen grains	subnormal female fertility
ms ₃₅	prophase to metaphase I; rarely telo. II	disintegration in first division	empty	

Data on inheritance are presented in the next table. The segregations for ms_{34} border on significance for a deficiency of recessives. Within the limits of the small populations, all other segregations fit monogenic expectations.

Mutant	Family number	Generation	+	ms	Total
ms_{30}	60L1069	F ₂	22	8	30
	61L1172	BC	12	17	29
ms_{33}	61L1175	F ₂	22	8	30
	61L1638	F ₂	104	36	140
	Total		126	44	170
	Expected			42.5 ± 6.1	
ms_{34}	61L1176	F ₂	25	4	29
	61L1177	F ₂	23	6	29
	61L1639	F ₂	90	21	111
	Total		138	31	169
	Expected			42.25 ± 6.0	
ms_{35}	61L1178	F ₂	25	5	30
	61L1639	F ₂	88	28	116
	Total		113	33	146
	Expected			36.5 ± 5.3	

Rick, C. M. New mutations
at old loci.

Since the last list was issued
(TGC 8:31) five new instances
have been discovered of indepen-

dent appearances of previously known mutants. In each instance the new mutant was found in material so completely unrelated to the original source that it is either impossible or highly unlikely that the same two deviants had a common origin. The allele tests prove a common locus although they do not prove that the new allele is necessarily identical with the original one.

ag (LA422). The entire population of an accession native to Cristobal Island, Galápagos completely lacked anthocyanin. In contrast to typical ag, no anthocyanin was observed at any stage of development. Test crosses with a₁ and aw yielded normal progeny, but the cross with ag produced 26 greenstem seedlings.

e. A broad-leaved line was received from Dr. H. B. Cordner of Oklahoma State in 1959. He had received it in turn from his brother-in-law, who had discovered it as a variant of the var. Big Boy growing at Orem, Utah. In the seedling stage it resembles both c and e, but in later development it resembles the latter to a greater extent by its waviness of the midvein, tendency of lower segments to web with the midvein, and other leaf distortions. Hybrids with c gave only normal seedlings, but the cross with an e stock gave 5 seedlings with undivided leaves of the e phenotype.

ff (2-505). In searching a field of V-San Marzano-A (a selected line not yet introduced) our attention was attracted to a partly fruitful plant with distinctive foliage. The leaf shape of this plant and of its progeny was remarkably similar to that of filicifolium. An allele cross between the

two lines yielded a progeny of 21 seedlings, all with ff phenotype. Numerous plants of the same phenotype, therefore likely allelic to ff, have been encountered subsequently in the same variety, suggesting that the new mutation occurred in an early generation of the seed increase of this new line.

l₁ (2-491). This variant was found as an off-type in the variety Roma by Mr. Melvin Zobel. According to his description and our experience with progeny of the original plant, foliage is yellow, more intense in older leaves, stigmas lack chlorophyll, and fruits have less pigment at all stages. Our suspicions that this might be an allele of l₁ were confirmed by the lutescent phenotype of all nine plants in the progeny of a test cross. The more extreme phenotype of the new mutant suggests that it might represent a different allele. This possibility will be tested with F_2 segregation.

ps (2-471). Discovered as a highly unfruitful plant in variety San Marzano, this mutant proved identical with the original ps in every aspect of floral structure. It bred true from the seeds that had been naturally set. With very few exceptions, it has been our experience that mutants that are identical in a specific phenotype turn out to be allelic. It is therefore very likely that in 2-471 we are dealing with another appearance of ps, adding San Marzano to the list of varieties--John Baer and Pearson--in which this mutation has spontaneously occurred.

Rick, C. M., and W. H. Dempsey
Trisomic segregations for cm.

Recent trisomic segregations are of interest in respect to the genes cpt and cm. Since data

for the former are given in the note concerning segregation and linkage relations of that gene, they need not be repeated here. Figures allegedly locating cm on chromosome 11 were presented in TGC 9:42. Difficulties were encountered in the classification of cm, and it was difficult to account for the appearance of a high proportion of cm/cm/cm triplo-11 individuals. The data for cm-triplo-11 are presented again herewith because an error was made in reporting them; an additional family is also reported for this combination. In the meanwhile data, also included below, have been secured showing significant deviations in the direction of trisomic segregation for cm in triplo-10.

Family	Chromosome	Phenotype	2N	2N+1	Total
57L78-80	11?	<u>cm</u> ⁺	209	73	282
		<u>cm</u>	32	4	36
58L285	11?	<u>cm</u> ⁺			130
	Seedling	<u>cm</u>			29
59L466	11?	<u>cm</u> ⁺	89	22	111
		<u>cm</u>	9	0	9
59L66	10	<u>cm</u> ⁺	67	26	93
		<u>cm</u>	14	0	14
59L464	10	<u>cm</u> ⁺	77	25	102
		<u>cm</u>	17	0	17

In the above figures only the family 59L466 gives a strong trisomic ratio, yet all families deviate significantly from normal inheritance and the absence of trisomic cm segregants is suggestive. It is interesting to note that simultaneous segregations in trisomics for other chromosomes tended to yield more than the expected proportion of cm (1958 data: 74+:50 cm/, 103+:51 cm/, 107+:69 cm/, 139+:80; 1959 data: 71+:29 cm). Heterozygous expression of cm

might account for such excessive frequencies of cm. Whatever the explanation, the control results contrast sharply with those for triplo-11? and triplo-10. Although the data are not unequivocal, they strongly suggest that cm was located on the extra chromosome in both trisomics.

The presence of cm on chromosome 10 and 11? fortifies the suspicions that the latter is not a primary trisomic. In many progenies of 11? we have also observed a fair frequency of triplo-10 and a few of triplo-7, suggesting that 11? might be a tertiary with parts of 7 and 10 constituting the extra chromosome. Until recently, however, we could not confirm this cytologically. In new smears made in 1961, however, this material reveals an occasional pentavalent. The rarity of such configurations might result from an unequal interchange between 7 and 10.

Evidence from radiation-induced deletions (Genetics 46:1231-1235, 1961) clearly established that chromosome 11 carries linkage group V. Since earlier trisomic tests between triplo-11? had not given trisomic ratios for genes of group V, triplo-11? cannot be triplo-11. All of this information is therefore compatible in establishing the linkage group for chromosome 11 and suggesting that triplo-11? is a tertiary trisomic with the extra being an interchange between 7 and 10. Bona fide triplo-11 is being sought in our material and a number of likely candidates in the progeny of triploids are being investigated.

Rick, C. M., and F. W. Martin
Continued linkage tests with
mutants of Stubbe's group I.

Linkage tests have been continued,
following the same procedures
reported in TGC 10 and 11. Two
new linkages have been encountered.

Results of all completed tests between these and an assortment of markers are given in the following table. Linkages are indicated by L, suggested but non-significant indications of linkage by S, and no significant departures from random recombination by X. In order to conserve space, data for the last two categories are not presented but records are kept for anyone who might want them. Of special interest in this tabulation is the linkage of flavescens with both y and pr. Since these results have been confirmed in several independent tests, pr and fla must be on chromosome 1, providing better seedling markers for that chromosome. The following tabulation also summarizes tests with six mutants of the Stubbe III series (see note by Hansen, Rick, and Boynton).

Linkages with Dr. Stubbe's mutants detected in 1961

Testers	Ch's'm	Mutants							
		Stubbe I		Stubbe III					
		def	fla	bip	gri	id	mu	oli	spa
a ₁	11	X	X	X	X	L	X	X	X
ah	9	X	X	X	X	X	X	X	X
au	1			X	S			X	
c	6	L	X	S	X	X	L	X	X
d ₁	2	X	X	L	X	X	S	X	X
d ₁	8	X	X	X	X	X	X	X	L
e	4	X	X		L		S	X	X
H	10		X	X	X			L	
l ₁	8	X	X		X		X	X	L

(cont. on next page)

Linkages with Dr. Stubbe's mutants detected in 1961 (cont.)

Testers	Ch's'm	Mutants							
		Stubbe I		Stubbe III					
		def	fla	bip	gri	id	mu	oli	spa
La	?	X	X	X		X	X	X	
pr	1?	X	L						
r	3		X						
rv	?		X						
sf	?		X						
tf	7		X	X	X			X	
w ₄	?		X						
W ^m	2				X				
y	1		L						
yv	6	L		X	X	X	X	X	

Segregation data for the linked combinations are presented below. Data for the fla-pr were given in TGC 11:23.

Combination	++	+t	m+	mt	Adj. cont. chi-square	co.
def - c	132	33	31	3	1.68	37.0
def - yv	47	22	21	0	7.22	0-20.0
	117	45	50	2	11.79	21.5
fla - y	22	9	8	1	0.43	
	Selected for y		26	1	5.44	
	46	23	15	1	3.47	24.0

Stocks of these mutants and pertinent segregating progenies have been transmitted to cooperators in charge of the respective chromosomes.

Smith, P. G. Linkage
with the Tm₂ gene.

Selfed populations of lines
supposedly having the linkage
between the Tm₂ and nv genes

broken (Clayberg, C. D., TGC 11) yielded 0 to 2 per cent virulent plants. None of these lines were homozygous Tm₂/Tm₂, but segregated in approximately a 2:1 ratio for TMV resistance.

It would appear that the presumed broken linkage is instead an added lethal factor, the cause of which has not been determined. Because of this, however, breeding programs using the presumed broken linkage lines should be reassessed in the light of the above information.

Snoad, B. Pachytene chromosomes
and the linkage maps.

A comparison of the genetic
activity of the euchromatin and
heterochromatin of tomato chromo-

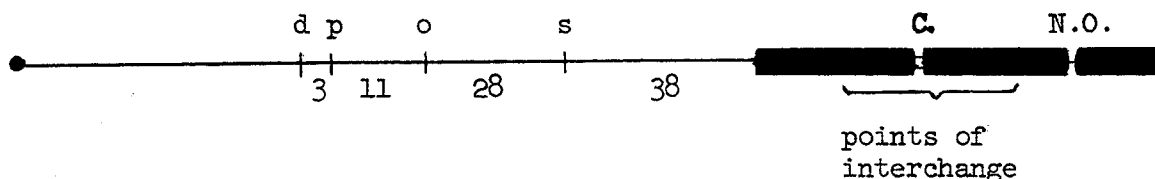
somes is being attempted by means of X-ray-induced interchanges. The chromosomes involved in each interchange are identified at pachytene, the point of interchange plotted and the linkage between this point and selected genes is then estimated.

Pollen treated with 4000r was used to produce 490 plants of which 24 were subsequently determined to have interchanges. In only two of these plants had an interchange taken place in the euchromatin.

Seven plants, each with an interchange in the heterochromatin of chromosome 2, have been used in test crosses. Linkage between d, p, o, s and the points of interchange was determined. The results show that there is no direct relationship between linkage values and the physical distance between a gene and a point of chromosome breakage in the heterochromatin. For example, the linkage value between T (the point of interchange) and s remained reasonably constant around 38 units although the physical distance varied considerably in these seven plants.

As crossing-over probably does not occur in the heterochromatin the linkage values must be assumed to measure the distance between the genes and the point on the chromosome at which euchromatin is replaced by heterochromatin.

Thus the map of chromosome 2, from my figures, appears to be:



An interchange between chromosomes 1 and 2 has brought about linkage between y (on chromosome 1) and s (on chromosome 2). It would seem that y is only 13 units from the heterochromatin in the long arm of chromosome 1 and it will be interesting to find out where br and Cf are situated in relation to the centromere considering that each is some 30 units either side of y.

It is hoped that other interchanges in the euchromatin will be isolated so that a more accurate idea of the relationship between a chromosome, as seen at pachytene, and the linkage map may be obtained. Meanwhile chromosomes other than number 2 will soon be under test.

Stettler, R. F. Dosage effect
of the lanceolate gene.

In a study investigating
morphogenetic effects of the La
gene ++ and La+ sister plants

of isogenic background were treated with colchicine. Suspected tetraploid shoots were subjected to pollen analysis and chromosome counts, and only verified inflorescences were selected for reciprocal crosses (LaLa++ x ++++). Excessive flower drop and a Fusarium outbreak reduced the number of successful crosses drastically. Two progenies with a total of 20 individuals were recovered and germinated. They were classified as 3 normal plants with compound leaves, 4 plants with simple leaves and 13 plants with clearly intermediate phenotype. The segregation ratio was fairly close to expectation on the basis of either chromosome or chromatid segregation. The third and later leaves of the 13 intermediate plants were compound with 2 lateral leaflets and a proportionally long terminal leaflet in contrast to the relatively short terminal and 4 laterals of the normal tetraploid (++++). The new segregants are, therefore, considered to be of La+++ genotype subject to verification by breeding tests. These findings do not agree with those reported by L. Monaco in TGC 11, 1961, according to which the La+++ genotype was supposed to have the characteristics of the normal tomato leaf.

Further evidence for the dosage effect of La was recently found when a triploid seedling, confirmed by chromosome counts, arose spontaneously in the progeny of a selfed La+ plant. Its leaves showed a range of variation between the phenotypes of LaLa++ and La+++, and it is, therefore, assumed to be La++. This hypothesis is now being tested.

Thompson, A. E. A new unstable chlorophyll mutant.

A new unstable chlorophyll mutant was found this year that resembles ghost (gh), but is not nearly as

extreme in its expression. The seed came from an amateur "Luther Burbank" residing near Chicago, and the original parentage of the mutant is unknown. Inheritance studies and tests for allelism with gh are in progress. It is very likely a recessive since plants of the F_1 between two normal lines and the mutant are normal in all respects. The cotyledons appear normal, but small albino sectors of a superficial nature may usually be detected on the first true leaves. The terminal growing point does not progress continually toward a more extreme expression of albinism as is usually observed with ghost. Both white and yellow phase albino sectors on leaves and stems have been observed. The albino sectors are well expressed in the greenhouse during the winter months, but surprisingly good vegetative growth is maintained. Normal or near normal male and female fertility has been observed. Flowers and fruits occasionally show a few light green sectors, but apparently normal development of carotenoid pigments occurs. Plants transplanted and grown in the field during the past summer were nearly normal in all respects. Small albino sectors were found only on the first true leaves. Classification of segregating material under field conditions may prove to be difficult. Ghost plants also tend to fare better during late spring, summer, and early fall thus extending the similarity between ghost and the new mutant.

Fruits from plants of the mutant grown in the field were analyzed for total carotenoid pigments and carotene in comparison with three normal varieties and one high-pigment line. Fruits were harvested at the turning stage and ripened for 14 days in an air-conditioned chamber at $65^\circ \pm 1.5^\circ\text{F}$. Measurements of total carotenoids and carotene were made using the spectrophotometric method of McCollum.

Variety	Total	Carotene	T/C	Hunter a/b
Acc. 387	5.54	0.46	12.04	1.86
Garden State	6.02	0.24	25.08	1.94
Kc 109	6.26	0.27	23.19	1.99
Kc 146	6.52	0.46	14.17	2.05
1252-102 (<u>hp/hp</u>)	12.76	0.58	22.00	2.48

The data indicate that the new mutant (Acc. 387) contains slightly less total carotenoid pigments than the normal varieties. At this time it is not known if the apparent reduction in total carotenoids is caused by the mutant character. Work is currently in progress to determine if higher than normal levels of colorless carotenoids are metabolized as is the case with ghost.

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PART IIIADDITIONS TO STOCK LIST

(Last complete stock list issued in TGC 10; see also additions in TGC 11 and sources in gene lists in this issue)

STOCKS AVAILABLE

<u>Source</u>	<u>Description</u>																																														
Denby, L. G.	Verticillium-resistant Ve Ve strains of the following established varieties. (Resistance from the variety Loran Blood, supplied by Dr. O. S. Cannon).																																														
	<table> <tr><td>Ace</td><td>Pennheart</td></tr> <tr><td>Bonny Best</td><td>Pritchard</td></tr> <tr><td>Bounty</td><td>Puck</td></tr> <tr><td>Break o' Day</td><td>Red Chief</td></tr> <tr><td>Clark's Early (Dohler #7)</td><td>Red Cloud</td></tr> <tr><td>Earliest of All</td><td>Red Jacket</td></tr> <tr><td>Early Baltimore</td><td>Rutgers</td></tr> <tr><td>Early Chatham</td><td>Signet</td></tr> <tr><td>Early Harkness</td><td>Sioux</td></tr> <tr><td>Early Jersey</td><td>Speed</td></tr> <tr><td>Early Lethbridge</td><td>Splendid</td></tr> <tr><td>Farthest North</td><td>Stokesdale #4</td></tr> <tr><td>Fireball</td><td>Sugawara</td></tr> <tr><td>Firesteel</td><td>Summerland Gem</td></tr> <tr><td>Geneva #6</td><td>Superior</td></tr> <tr><td>Harris Gem</td><td>Valiant</td></tr> <tr><td>John Baer</td><td>Valnorth</td></tr> <tr><td>Longred</td><td>Victor</td></tr> <tr><td>Manasota</td><td>Wasatch</td></tr> <tr><td>Marmande</td><td>Windowbox</td></tr> <tr><td>Meteor</td><td>Wisconsin 55</td></tr> <tr><td>Morses 498</td><td>Wisconsin (James)</td></tr> <tr><td>Non-acid</td><td></td></tr> </table>	Ace	Pennheart	Bonny Best	Pritchard	Bounty	Puck	Break o' Day	Red Chief	Clark's Early (Dohler #7)	Red Cloud	Earliest of All	Red Jacket	Early Baltimore	Rutgers	Early Chatham	Signet	Early Harkness	Sioux	Early Jersey	Speed	Early Lethbridge	Splendid	Farthest North	Stokesdale #4	Fireball	Sugawara	Firesteel	Summerland Gem	Geneva #6	Superior	Harris Gem	Valiant	John Baer	Valnorth	Longred	Victor	Manasota	Wasatch	Marmande	Windowbox	Meteor	Wisconsin 55	Morses 498	Wisconsin (James)	Non-acid	
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PART VFINANCIAL STATEMENT

(to December 31, 1961)

		<u>Total</u>
<u>Balance from 1960</u>		\$161.38
<u>Receipts</u>		
Assessments	\$261.75	
Sale of back numbers	91.00	
Interest on savings	5.42	358.17
<u>Assets</u>		519.55
<u>Expenditures</u>		
TGC Report No. 11, 1961		
Multilithing and covers	155.49	
Stencils, envelopes, and clasps	17.87	
Postage	25.81	
Miscellaneous		
Postage for meeting notice	10.68	
Postage for newsletter	11.00	
Invoice form	.54	221.39
<u>Balance</u>		298.16

MEMBERSHIP STATUS

Assessments paid for 1961	38
1962	36
1963	149
1964	12
1965	2
1966	2
1968	1
1970	1
Total members	241